

Caloric Restriction Mimetics against Age-Associated Disease: Targets, Mechanisms, and Therapeutic Potential

Frank Madeo,^{1,2,*} Didac Carmona-Gutierrez,¹ Sebastian J. Hofer,¹ and Guido Kroemer^{3,4,5,6,7,8,9,*}

¹Institute of Molecular Biosciences, University of Graz, Graz, Austria

²BioTechMed Graz, Graz, Austria

³Equipe 11 labellisée Ligue contre le Cancer, Centre de Recherche des Cordeliers, INSERM U 1138, Paris, France

⁴Metabolomics and Cell Biology Platforms, Gustave Roussy Comprehensive Cancer Center, Villejuif, France

⁵Université Paris Descartes, Sorbonne Paris Cité, Paris, France

⁶Université Pierre et Marie Curie, Paris, France

⁷Pôle de Biologie, Hôpital Européen Georges Pompidou, Paris, France

⁸Karolinska Institute, Department of Women's and Children's Health, Karolinska University Hospital, Stockholm, Sweden

⁹Center of Systems Medicine, Chinese Academy of Science, Suzhou, China

*Correspondence: frank.madeo@uni-graz.at (F.M.), kroemer@orange.fr (G.K.)

<https://doi.org/10.1016/j.cmet.2019.01.018>

The increase in life expectancy has boosted the incidence of age-related pathologies beyond social and economic sustainability. Consequently, there is an urgent need for interventions that revert or at least prevent the pathogenic age-associated deterioration. The permanent or periodic reduction of calorie intake without malnutrition (caloric restriction and fasting) is the only strategy that reliably extends healthspan in mammals including non-human primates. However, the strict and life-long compliance with these regimens is difficult, which has promoted the emergence of caloric restriction mimetics (CRMs). We define CRMs as compounds that ignite the protective pathways of caloric restriction by promoting autophagy, a cytoplasmic recycling mechanism, via a reduction in protein acetylation. Here, we describe the current knowledge on molecular, cellular, and organismal effects of known and putative CRMs in mice and humans. We anticipate that CRMs will become part of the pharmacological armamentarium against aging and age-related cardiovascular, neurodegenerative, and malignant diseases.

Caloric Restriction Improves Health

Caloric restriction (CR) consists of the chronic reduction of total calorie intake without malnutrition. Together with intermittent fasting (which can be regarded as a particular form of CR in which episodes of *ad libitum* feeding are alternated with episodes of up to zero caloric uptake), CR is the only known strategy to robustly improve health- and lifespan in most, if not all, living organisms. In Rhesus monkeys, two differently designed studies revealed contrasting results on lifespan (Mattison et al., 2017) but similar health benefits and delayed onset of aging phenotypes. In humans, CR has been reported to counteract several age-associated alterations (Figure 1). In non-obese, healthy adults, 24 months of continuous CR (15%–25%) was safe (Romashkan et al., 2016), improved the quality of life (Martin et al., 2016), and caused 10%–13% weight loss (mostly, but not exclusively, reducing fat mass), which stabilized after 1 year (Redman et al., 2018). Fasting insulin levels, body temperature (a possible marker for metabolic rate), resting energy expenditure, oxidative stress, and thyroid axis activity were reduced under CR (Il'yasova et al., 2018; Redman et al., 2018). “Metabolic adaptation,” a long-term effect of CR that reduces the metabolic rate below the expected value, occurs in humans and may support longevity (Heilbronn et al., 2006; Redman et al., 2018). In healthy humans, CR also decreases the levels of circulating tumor necrosis factor- α and cardiometabolic risk factors (triglycerides, cholesterol, and blood pressure) (Most

et al., 2018; Ravussin et al., 2015). Upon CR and weight loss, insulin growth factor-1 (IGF1) levels and insulin resistance are reduced in obese patients (Dubé et al., 2011). However, they are not improved in non-obese humans after the 1-year weight loss phase (Most et al., 2018) (contrary to mouse studies) unless protein intake is also reduced (Fontana et al., 2008). While CR inhibits inflammation, its effects on immunity need further clarification since different levels of CR may subvert and/or modulate immune defenses against bacterial (Tang et al., 2016) and viral infection (Wang et al., 2016). In obese humans, CR promotes significant weight loss and improves general health (Ard et al., 2017). Of note, the well-documented good health and high incidence of centenarians in the population of the Japanese Okinawa island have been attributed to nutritional cues including a mild and consistent CR (~10%–15%) (Willcox and Willcox, 2014).

Molecular Effects of CR and Fasting

Macroautophagy (hereafter referred to as autophagy) is a conserved cellular recycling program that eliminates dysfunctional organelles, proteins, and aggregates from the cytoplasm, hence protecting cellular functionality and integrity. Accordingly, impaired or dysregulated autophagy has been linked to advanced age, neurodegeneration, cardiovascular diseases (CVDs), and cancer. In turn, the activation of autophagy via genetic or pharmacological means extends lifespan and/or



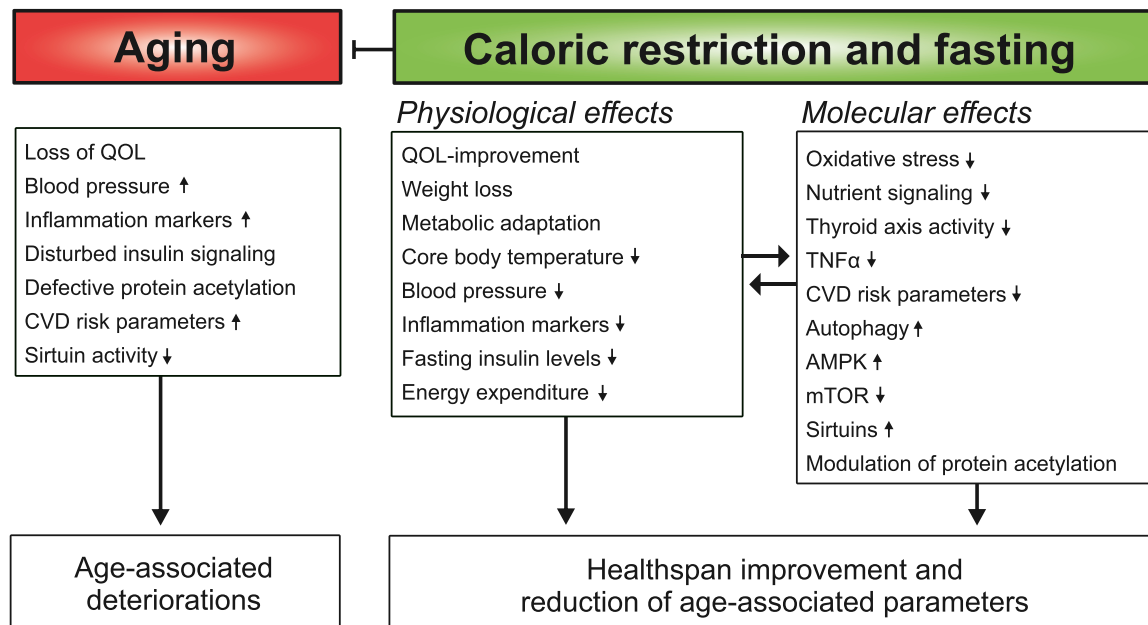


Figure 1. Physiological and Molecular Effects of Caloric Restriction

Caloric restriction reverses aging-derived effects by igniting numerous pathways involved in the improvement of health parameters. AMPK, AMP-activated protein kinase; CVD, cardiovascular disease; mTOR, mechanistic target of rapamycin; TNF α , tumor necrosis factor alpha; QOL, quality of life.

healthspan in numerous model organisms, including mice (Eisenberg et al., 2009). As a catabolic process, autophagy is induced upon nutrient deprivation and plays an important role in the beneficial effects exerted by CR and fasting regimens. CR modulates several molecular key players involved in the regulation and execution of autophagy, nutrient signaling, and energy metabolism (Figure 1). For instance, CR activates AMP-activated protein kinase (AMPK) (Cantó and Auwerx, 2011). AMPK is an energy sensor that inhibits the kinase activity of mechanistic target of rapamycin (mTOR), an autophagy repressor, under CR. Furthermore, CR directly and indirectly activates sirtuins (SIRT), which are nicotinic adenine dinucleotide (NAD⁺)-dependent lysine deacetylases (KDACs) and play central roles during aging and autophagy (Guarente, 2007). SIRT1 and AMPK may engage in a positive feedforward loop to amplify the response to CR.

Protein acetylation is a major regulator of autophagy. The N ϵ -acetylation of lysines is a phylogenetically conserved, post-translational protein modification that is catalyzed by lysine acetyltransferases (KATs) and reversed by KDACs. N ϵ -acetylation regulates multiple metabolic enzymes, facilitating the adaptation to nutrient availability. Of note, N ϵ -acetylation may occur in a non-enzymatic fashion in the presence of AcCoA, especially at an acidic pH (James et al., 2017). There are four ways to diminish N ϵ -acetylation of proteins: (1) by reducing the concentration of cytosolic AcCoA, the sole donor of acetyl groups used by KATs, e.g., via inhibition of its synthesis from glycolysis, β -oxidation of fatty acids, or the catabolism of branched amino acids, or via increase of its consumption, for instance by carnitine acetyltransferases that transfer AcCoA acetyl groups on carnitine; (2) by degrading S-acetyl glutathione by mitochondrial thioesterase glyoxalase 2, GLO2, or cytosolic GLO1, thus reducing intermedi-

ates for non-enzymatic N ϵ -acetylation; (3) by activating specific KDACs, mostly SIRT; and (4) by inhibiting KATs such as E1A-binding protein p300 (EP300). Notably, SIRT1 activity is low in aged and obese mice. This correlates with the inhibitory hyperacetylation of SIRT3, and transgenic activation of SIRT3 may improve the hepatic consequences of obesity including glucose intolerance (Kwon et al., 2017). Moreover, in mice, transgene-enforced overexpression of SIRT6 (Kanfi et al., 2012) or brain-specific expression of SIRT1 (Satoh et al., 2013) is sufficient to extend lifespan. In an earlier study, however, whole-body overexpression of SIRT1 did not extend lifespan (Herranz et al., 2010). Similarly, another report observed no lifespan extension upon overexpression of the SIRTs sir-2.1 and dSir2 in the nematode *Caenorhabditis elegans* and the fruit fly *Drosophila melanogaster*, respectively (Burnett et al., 2011), thus contradicting previous results (Bauer et al., 2009; Rogina et al., 2002; Tissenbaum and Guarente, 2001). While it seems clear that SIRTs exert important functions related to healthy aging, their specific role in promoting longevity remains to be clarified (Dang, 2014).

Interestingly, autophagy and protein acetylation are subjected to circadian fluctuations (Sato et al., 2017). This oscillation is lost with aging and has been proposed as a modulatory target of CR (Sato et al., 2017). The maintenance of rhythmic (de)acetylation by CR is hypothetically linked to increased NAD⁺ levels, coupled to SIRT1 activation and rhythmic changes in the inhibitory acetylation of acetyl-CoA-generating acyl-CoA synthase short-chain family member 1 (ACSS1) (Sato et al., 2017). In aged flies, protein acetylation is increased, a phenomenon that can be attenuated by reducing the AcCoA-generating enzyme ATP citrate lyase (ACLY) or by mutating the KAT Chameau, resulting in an extended lifespan (Peleg et al., 2016). Similarly, the inhibitory

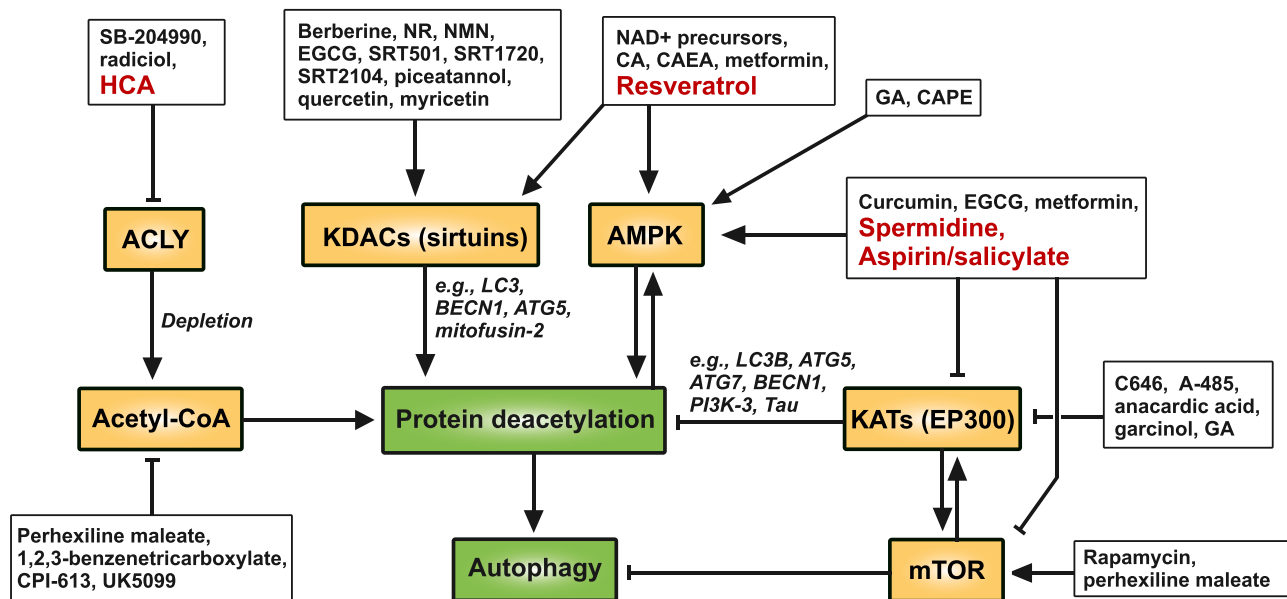


Figure 2. Mechanistic Targets of Known and Potential CRMs

The displayed compounds (known CRMs shown in red, potential CRMs in black) converge in protein deacetylation via acetyl-CoA depletion, inhibition of acetyltransferases, or stimulation of deacetylases, ultimately resulting in autophagy activation. Moreover, AMP-activated protein kinase (AMPK) and mechanistic target of rapamycin (mTOR) represent further targets of CRMs, the pro-autophagic activity of which is intertwined with protein deacetylation processes. ACLY, ATP citrate lyase; BECN1, Beclin 1; CA, caffeic acid; CAEA, caffeic acid ethanalamide; CAPE, caffeic acid phenyl ester; EGCG, Epigallocatechin-3-gallate; GA, gallic acid; HCA, hydroxycitric acid; KATs, lysine acetyltransferases; KDACs, lysine deacetylases; NMN, nicotinamide mononucleotide; NR, nicotinamide riboside; PI3K-3, phosphatidylinositol 3-kinase catalytic subunit type 3.

hyperacetylation of the pro-autophagic transcriptional factor Foxo1 has been observed in aged mouse hearts (Ren et al., 2017). Moreover, CR deacetylates histones H3 and H4 in mouse fat pads (Xu et al., 2015) and reduces the levels of biotin, which acts as an endogenous inhibitor of SIRT1 (Xu et al., 2015). Both histone deacetylation and deacetylation of cytosolic proteins may affect the expression and activity, respectively, of autophagy-relevant proteins (Eisenberg et al., 2009; Mariño et al., 2014). Both in mice and in humans, acute starvation causes a reduction of the acetylation of cytoplasmic proteins in peripheral blood mononuclear cells (Pietrocola et al., 2017). However, every-other-day fasting increases histone acetylation in the mouse retina (Guo et al., 2016), and acetylation is reduced in aged mouse livers, a phenomenon that is reversed by CR, which causes hepatic protein hyperacetylation (Sato et al., 2017). This is at odds with chronic alcohol abuse, which leads to NAD⁺ depletion and SIRT inhibition, resulting in hyperacetylation of multiple proteins in the liver (such as AMPK, β -catenin, histone H3, and the transcription factors SREBB2, PPAR α , FOXO1, NF κ B, and NFAT) (French, 2016). Therefore, the impact of CR on acetylation might depend on tissue, cell type, and the precise protein species. Indeed, one study reports that CR causes hyperacetylation of mitochondrial proteins in the liver and reduces acetylation in brown adipose tissue, yet it fails to affect the acetylation of mitochondrial proteins from other tissues (Schwer et al., 2009).

CR Mimetics

Despite the uncontestable health-promoting effects of CR, most individuals are unable to observe a CR lifestyle, likely explaining

some failures in observational clinical studies (Redman et al., 2018). Although long-term compliance may be improved by periodic fasting regimens, pharmacological approaches that induce autophagy without the subjective discomfort linked to CR or periodic fasting are warranted. Indeed, several CR mimetics (CRMs) improve health parameters in rodents and humans (see below). We previously defined CRMs as compounds that activate autophagy by promoting the deacetylation of cellular proteins (Madeo et al., 2014), by (1) depleting AcCoA, (2) inhibiting acetyltransferases, and/or (3) stimulating deacetylases (Figure 2).

This definition reflects the fact that protein acetylation usually inhibits autophagy, while protein deacetylation favors autophagy. For instance, starvation is coupled to the inhibition of the acetyltransferase EP300 (due to the depletion of AcCoA), as well as to the activation of the deacetylase SIRT1 (due to the increase of the NAD⁺/NADH ratio and the activation of AMPK). This results in the deacetylation of hundreds of cellular proteins (Morselli et al., 2011), reflecting multipronged regulatory effects on cell metabolism and the autophagic cascade. A systematic screen for KATs, the inhibition of which would induce autophagy, led to the identification of EP300 as a major negative regulator of autophagy that acts epistatic to starvation (Mariño et al., 2014).

Interestingly, EP300 is subjected to activating phosphorylation by mTORC1 (Wan et al., 2017), while conversely, inhibition of EP300 generally results in mTORC1 inhibition (Pietrocola et al., 2015), suggesting that both regulatory systems are intertwined. Similarly, protein deacetylation may be connected to the activation of AMPK, a potent autophagy inducer. Thus, deacetylation

of liver kinase B1 (LKB1), for instance by SIRT2, favors the LKB1-mediated activation of AMPK (Tang et al., 2017). Likewise, EP300 inhibition results in AMPK activation (Pietrocola et al., 2015). These examples illustrate how protein deacetylation may initiate autophagy, correlating with mTORC1 inhibition and AMPK activation. However, EP300 inhibition results in the induction of autophagy even in conditions in which AMPK is deleted, mTORC1 is artificially activated, or ULK1 is inhibited (Pietrocola et al., 2018; Su et al., 2017). This suggests that protein deacetylation can set off the autophagic cascade in a dominant fashion that is largely independent of other regulatory systems.

In accord with this interpretation, EP300 inhibition or SIRT activation may favor autophagy through deacetylation reactions that affect multiple autophagy-executory proteins (Pietrocola et al., 2015). For instance, EP300 inhibition results in the deacetylation of phosphatidylinositol 3-kinase catalytic subunit type 3 (PI3K3) at K29 and K771, favoring its interaction with allosteric activators contained in the pro-autophagic Beclin 1 (BECN1) complex and its substrate phosphatidylinositol, respectively (Su et al., 2017). BECN1 itself is also a substrate of EP300 and SIRT1 (at K430 and K437), and deacetylation of BECN1 favors the dissociation of its inhibitory interactor Rubicon (Sun et al., 2015). Of note, pro-autophagic derepression of BECN1 has been recently shown to promote longevity in mice (Mariño et al., 2014). Furthermore, SIRT1 deacetylates nuclear microtubule-associated proteins 1A/1B light chain 3B (hereafter referred to as LC3) (at K49 and K51), stimulating its interaction with the nuclear protein DOR and its export to the cytoplasm, where it acts as a key initiator of autophagy (Huang et al., 2015). EP300 can also acetylate ATG5 and ATG7, both of which are involved in a conjugation system that promotes LC3 lipidation, which is required for autophagy induction. Of note, ATG5 is also deacetylated by SIRT2, supporting the notion that many autophagy regulators are substrates of both EP300 and SIRTs (Liu et al., 2017a).

While the link between deacetylation of cytoplasmic proteins and autophagy seems rather unambiguous, it appears less straightforward with respect to nuclear proteins. On the one hand, the pro-autophagic transcriptional response has been linked, for example, to SIRT1- and spermidine-induced deacetylation of histones H4 and H3 (Eisenberg et al., 2009), respectively. On the other hand, glucose deprivation stimulates AMPK activation with the final result that acetyl-CoA synthetase 2 (ACSS2) phosphorylated by AMPK translocates to the nucleus where it interacts with the transcription factor EB (TFEB) and binds to promoter regions of autophagy genes, locally producing acetyl-CoA and favoring pro-autophagic H3 hyperacetylation (Li et al., 2017b). These divergent outcomes may reflect feedback loops that impose a self-limitation on the autophagic process. For instance, rapamycin-induced autophagy is coupled to the hypoacetylation of H4K16 following the downregulation of lysine acetyltransferase 8 (KAT8), thereby reducing the transcription of pro-autophagic genes (Füllgrabe et al., 2013).

Besides autophagy-regulatory and executory proteins, deacetylation may also affect autophagic substrates. Depletion of general control of amino acid synthesis 5 (GCN5) like-1 (GCN5L1), a component of the mitochondrial acetyltransferase machinery, leads to mitochondrial protein deacetylation catalyzed by SIRT3, thus favoring mitophagy (Webster et al., 2013).

SIRT1 deacetylates mitofusin-2, a protein tethered to the mitochondrial membrane, facilitating SIRT1-induced autophagy and mitophagy (Biel et al., 2016). As a further example, EP300 inhibition reduces the acetylation of Tau (a protein that forms pathogenic intraneuronal aggregates in Alzheimer's disease), which favors its clearance by autophagy (Min et al., 2015).

Bona fide CRMs and Candidate CRMs

Several agents may be considered as CRMs since they cause protein deacetylation deriving in autophagy induction (Figure 2). We suggest that CRMs should also have the capacity to reproducibly extend lifespan and/or healthspan in model organisms, hence extending the functional definition of CRMs by another criterion. Here, we enumerate compounds that either fully comply with these stringent criteria (*bona fide* CRMs) or that do so at least partially according to the current state-of-the-art (potential CRMs) (Table 1).

Resveratrol and Other SIRT1 Activators

Resveratrol is a polyphenolic phytoalexin that is particularly abundant in the skin of grapes and in red wine. It has been shown to promote longevity across species and to improve age-related parameters in mice. However, resveratrol seems to only prolong the lifespan of mice on a high-fat diet (HFD) (Baur et al., 2006), but not on regular chow. Still, resveratrol exerts a number of protective effects in mammalian models of metabolic syndrome, type 2 diabetes (an effect that is enhanced when resveratrol is combined with metformin), cancer, neurodegeneration, and CVD (Rajman et al., 2018). However, contrary findings have been reported recently on its efficacy against metabolic syndrome (Kjær et al., 2017). Interestingly, resveratrol can counteract the reduction of duodenal SIRT1 levels in rats fed an HFD, which is accompanied by improved insulin sensitivity (Côté et al., 2015). This indicates the potential of resveratrol as an agent to counteract obesity- and diabetes-induced insulin resistance as well as dysregulated glucose homeostasis. Moreover, resveratrol induces a CR-like transcriptional signature in mice and recapitulates metabolic changes of CR in humans (Timmers et al., 2011).

Several studies have examined resveratrol on primates, also showing SIRT1 induction, NF- κ B repression, improved insulin signaling, and attenuated inflammation in adipose tissue of high-fat, high-sugar (HFS)-fed animals (Rajman et al., 2018), coupled to reduced CVD risk parameters induced by HFS (Mattison et al., 2014). A large number of clinical trials assessing its effects on cancer, diabetes, obesity, non-alcoholic fatty liver (NAFL), neurological disease, and CVDs have been performed with mostly beneficial outcomes.

Resveratrol targets a number of stress-related cellular components, including AMPK (Rajman et al., 2018), which might represent a major molecular target, and the NAD⁺-dependent deacetylase SIRT1. Both AMPK and SIRT1 have been shown to be required for resveratrol-induced health promotion (Lagouge et al., 2006; Price et al., 2012). Resveratrol can stimulate SIRT1 (possibly indirectly), resulting in general protein deacetylation and autophagy induction (Morselli et al., 2010, 2011; Pietrocola et al., 2012).

Although a *bona fide* CRM, resveratrol is afflicted by rather low systemic availability and absorption. One strategy to improve

Table 1. Classification of Protective Substances as CRMs, Potential CRMs, and Other Compounds

Group	Substance	(Major) Molecular Target(s)	(Nutritional) Sources	Clinical Trials ^a
CRMs	aspirin (and salicylate)	AMPK, EP300, COX-1, COX-2, mTOR, NF-κB	willow bark, synthetic	several meta-analyses available, e.g., Cuzick et al., 2015 ; Raju et al., 2016
	hydroxycitric acid	ACLY	diverse tropical plants, <i>Garcinia cambogia</i> , and <i>Hibiscus sabdariffa</i>	meta-analyses on weight loss through HCA in Onakpoya et al., 2011 ; NCT00699413 and NCT01238887
	resveratrol	KDACs (SIRT1), AMPK, NF-κB	fruits, plants, and skin of grapes	reviewed in Berman et al., 2017 , e.g., NCT02621554
	spermidine	KATs (EP300), mTOR, AMPK	wheat germs, soybeans, and nuts	safety evaluation in Schwarz et al., 2018 ; neuroprotection (Wirth et al., 2018); NCT02755246, NCT03378843, and NCT03094546
Potential CRMs	1,2,3-benzenetricarboxylate	citrate transport protein	synthetic	–
	acipimox	niacin receptor 1	synthetic	several studies on obesity and diabetes, e.g., NCT00549614, NCT01488409, NCT00943059, and NCT01816165
	berberine	SIRT1	<i>Berberis vulgaris</i> and several other plants (roots and bark)	numerous phase 3 and 4 studies; reviewed in Imenshahidi and Hosseinzadeh, 2016
	caffeic acid	AMPK and sirtuins	eucalyptus bark	7 studies registered, e.g., NCT03070262; the clinical potential of CAPE reviewed in Murtaza et al., 2014
	catechin	pleiotropic, exact mechanism unknown	cocoa, tea, and red wine	mainly green-tea combinations tested, few single compound studies; reviewed in Chacko et al., 2010 ; e.g., NCT03213340, NCT00233935, and NCT00448513
	curcumin	AMPK, mTOR, and EP300	<i>Curcuma longa</i>	numerous including phase 3 and 4 studies; reviewed in Gupta et al., 2013 ; e.g., NCT03085680, NCT01052025, NCT01975363, and NCT00099710
	epicatechin	pleiotropic, exact mechanism unknown	cocoa, tea, and red wine	numerous studies using catechin-rich extracts, few single component trials, e.g., NCT01856868, NCT01880866, NCT02221791, NCT01691404, NCT02490527, and NCT02292342
	EGCG	AMPK, mTOR, HATs, and KDACs	green tea	numerous phase 3 and 4 studies; reviewed and discussed in Mereles and Hunstein, 2011
	gallic acid	AMPK and HATs	black tea and various plants	mainly polyphenolic combinations tested, e.g., NCT02800967, NCT02005939, and NCT03214276
	metformin	AMPK, mTOR, HATs, and KDACs (sirtuins)	French lilac (<i>Galega officinalis</i>)	numerous phase 3 and 4 studies, reviewed in Nasri and Rafeian-Kopaei, 2014 ; e.g., NCT02432287
	myricetin	SIRT1	black tea, cole, parsley, garlic, curcuma, and fruits	reviewed in Li and Ding, 2012
	NAD ⁺	KDACs (sirtuins), AMPK	various food	reviewed in Fang et al., 2017 ; many studies supplementing precursors, especially NR
	nicotinamide	KDACs (sirtuins)	various food	numerous, e.g., NCT02213094, NCT02416739, NCT03061474, and NCT01250990
	nicotinamide mononucleotide	KDACs (sirtuins)	various food	NCT03151239 and UMIN000021309 (NIPH, Japan)
	nicotinamide riboside	KDACs (sirtuins)	various food	numerous studies, including phase 3 and 4; reviewed in Rolfe, 2014 , e.g., NCT03423342, NCT03423342, and NCT02921659
	perhexiline maleate	carnitine O-palmitoyl transferase 1, mTOR	synthetic	numerous; reviewed in Chong et al., 2016
	piceatannol	SIRT1	passion fruit seeds	–
	quercetin	SIRT1	black tea, onions, rocket, cole, curcuma, and fruits	reviewed in Miles et al., 2014 ; e.g., NCT00065676 and NCT01691404
	rapamycin	mTOR	<i>Streptomyces hygroscopicus</i>	numerous; reviewed in Li et al., 2014 , e.g., NCT01649960
	SRT1720	SIRT1	synthetic	–
	UK5099	mitochondrial pyruvate carrier	synthetic	–

(Continued on next page)

Table 1. Continued

Group	Substance	(Major) Molecular Target(s)	(Nutritional) Sources	Clinical Trials ^a
Others	4,4'-dimethoxychalcone	GATA transcription factors	<i>Angelica keiskei</i>	–
	A-485	EP300	synthetic	–
	acarbose	α -glucosidase	bacterial (<i>Streptomyces</i> , <i>Actinoplanes</i>)	numerous including phase 3 and 4 studies; e.g., NCT02865499, NCT02953093, and NCT01490918
	anacardic acid	EP300	cashew nutshell, <i>Anacardium occidentale</i>	–
	C646	EP300	synthetic	–
	CAEA	AMPK, sirtuins	synthetic	–
	CAPE	AMPK	propolis	clinical potential reviewed in Murtaza et al., 2014
	CPI-613	pyruvate dehydrogenase	synthetic	several phase 2 studies; e.g., NCT01835041, NCT03370159, and NCT01902381
	garcinol	EP300	<i>Garcinia indica</i>	–
	glucosamine	hexokinase and mTOR	crustaceans, cartilage	numerous studies on arthritis, reviewed in Ogata et al., 2018 ; e.g., NCT02448199
	radicol	ACLY and HSP90	<i>Monosporium bonorden</i>	–
	SRT501	SIRT1	see resveratrol	reviewed in Berman et al., 2017
	SB-204990	ACLY	synthetic	–
	SRT2104	SIRT1	synthetic	several phase 1 studies; three phase 2 studies registered: NCT01018017, NCT01154101, and NCT01018017

Classification was based on whether compounds are known (1) to induce protein deacetylation that is causal for protective autophagy and to exert health-promoting effects in higher models (CRMs), (2) to promote protective autophagy and have molecular targets involved in protein deacetylation (potential CRMs), and (3) to exert protective effects without evidence for either autophagy induction or protein deacetylation (others). ACLY, ATP citrate lyase; AMPK, AMP-activated protein kinase; CAEA, caffeic acid ethanolamide; CAPE, caffeic acid phenyl ester; COX, cyclooxygenase; EGCG, epigallocatechin-3-gallate; EP300, E1A-binding protein p300; HCA, hydroxycitric acid; HSP90, heat shock protein 90; KATs, lysine acetyltransferases; KDACs, lysine deacetylases; mTOR, mechanistic target of rapamycin; NF- κ B, nuclear factor “kappa-light-chain-enhancer” of activated B-cells; SIRT1, sirtuin (silent mating type information regulation 2 homolog) 1.

^aIf applicable, reviews and meta-analyses, or examples of advanced clinical studies, are depicted, indicating the current average phase of trials; clinicaltrials.gov identifiers, if not stated otherwise

this galenic problem consists in micronization to decreased particle size, yielding the proprietary formulation SRT501.

Other small-molecule activators of SIRT1 have been developed. For instance, SRT1720 has been demonstrated to extend lifespan and improve metabolic syndrome, insulin sensitivity, and endothelial dysfunction in mice ([Hubbard and Sinclair, 2014](#)). A related compound, SRT2104, which also extends murine lifespan, has undergone clinical phase I and II trials, revealing only minor adverse effects ([Hubbard and Sinclair, 2014](#)). Both SIRT1 activators have been shown to improve healthspan in mice, reducing inflammation and protecting from neurodegeneration ([Hubbard and Sinclair, 2014](#)). According to one clinical study, SRT2104 can reduce the serum levels of interleukin-6 and C-reactive protein induced by intravenous injection of lipopolysaccharide ([van der Meer et al., 2015](#)). Additional data on SRT2104 effects on human health will likely be reported in the near future.

Spermidine

Spermidine is a polyamine that induces autophagy in different model organisms, including mice ([Eisenberg et al., 2009, 2016](#); [Morselli et al., 2011](#)), and this induction is causal for at least some of the observed beneficial effects. For instance, genetic ablation of autophagy abrogates spermidine-mediated lifespan extension in yeast, nematodes, and flies and attenuates cardio-

protective effects ([Eisenberg et al., 2016](#)) in mice. Spermidine inhibits the activity of several acetyltransferases ([Eisenberg et al., 2009](#)), including EP300, and this suffices for autophagy induction ([Pietrocola et al., 2015](#)). Intriguingly, these pro-autophagic deacetylation effects are synergistic with those of resveratrol ([Morselli et al., 2011](#)), which instead promotes the deacetylase activity of SIRT1 (see above). Moreover, spermidine has been shown to inhibit mTORC1 and activate AMPK ([Mariño et al., 2014](#)). It has also been speculated that spermidine might post-translationally hypusinate the translation factor eIF5A, which leads to the synthesis of the pro-autophagy transcription factor TFEB, at least in immune cells ([Zhang et al., 2018](#)). Moreover, spermidine can promote mitophagy (a specialized form of autophagy that eliminates damaged or dysfunctional mitochondria) in cell culture ([Qi et al., 2016](#)) and mice ([Eisenberg et al., 2016](#)). In human cells, this depends on ataxia-telangiectasia mutated protein kinase (ATM) and consequently on the phosphatase and tensin homolog (PTEN)-induced putative kinase 1 (PINK1) ([Qi et al., 2016](#)), which has been linked to the promotion of mitophagy ([Eiyama and Okamoto, 2015](#)).

Spermidine is naturally produced in the body by cellular biosynthesis as well as by the intestinal microbiota. In addition, oral ingestion of spermidine contained in food items like wheat germs, soybeans, or nuts among others, results in a

good bioavailability (Soda et al., 2009). Besides its role in general cell homeostasis (e.g., stabilization of DNA and RNA, cell growth, and translation regulation), dietary supplementation of spermidine has been associated to manifold health-promoting effects (Madeo et al., 2018). Spermidine feeding promotes lifespan across species, including mice, suppresses tumorigenesis, enhances anticancer immune response, stimulates memory T cell formation, promotes cardioprotection, improves skeletal muscle regeneration, and mediates neuroprotection (Madeo et al., 2018), thus qualifying this polyamine as a CRM. Accordingly, increased levels of whole-blood spermidine are linked to longevity in healthy nonagenarians and centenarians (Pucciarelli et al., 2012). Furthermore, epidemiological studies correlate elevated dietary polyamine uptake with diminished cardiovascular and cancer-related mortality (Eisenberg et al., 2016). That said, increased polyamine levels have also been associated with various human pathologies (Madeo et al., 2018), although this might represent a (non-causal) protective response.

Clinical trials are needed to explore possible contraindications of spermidine administration. A trial on supplementation of spermidine-rich plant extracts to elderly is currently ongoing (www.clinicaltrials.gov identifier: NCT02755246) and suggests good tolerability and safety of the compound (Schwarz et al., 2018). Moreover, a small pilot trial has already revealed the beneficial effects of spermidine supplementation in elderly people with subjective cognitive decline (Wirth et al., 2018).

Hydroxycitric Acid and Other AcCoA-Depleting Agents

Hydroxycitric acid (HCA) acts as a competitive low-affinity inhibitor of ATP citrate lyase (ACLY), which generates cytosolic AcCoA and thus represents an AcCoA-depleting CRM. HCA is present in diverse tropical plants, including *Garcinia cambogia* and *Hibiscus sabdariffa*. HCA salts have been shown to reduce body weight, insulin resistance, and oxidative stress in obese Zucker rats (Asghar et al., 2007). In a mouse model of multiple sclerosis, a garcinia extract containing 50% HCA exerted anti-inflammatory and anti-oxidative effects (Goudarzvand et al., 2016). Furthermore, in mice, HCA improves the antitumor efficacy of immunogenic chemotherapy, which required regulatory depletion of T cells (which dampen anticancer immunity) from the tumor bed (Mariño et al., 2014; Pietrocola et al., 2016) and tumors to be autophagy-competent (Mariño et al., 2014). Indeed, HCA promotes autophagic flux in diverse organs, including the liver, the myocardium, and skeletal muscle, and this induction is required for body weight reduction in mice (Mariño et al., 2014). HCA is an over-the-counter weight-loss drug, and clinical trials have shown its effectivity in obese patients, although only at high doses (≥ 3 g per day). Nevertheless, several rodent studies showing adverse effects on the male reproductive system upon administration of HCA preparations have incited health concerns.

Another ACLY inhibitor, the synthetic SB-204990, also stimulates autophagy in mice (Pietrocola et al., 2016) and mediates cholesterol and triglyceride reduction as well as tumor growth suppression in rodents (Pietrocola et al., 2016). Possibly, the Cullin3-KLHL25 (Kelch-like family member 25) ubiquitin ligase is responsible for the degradation of ACLY and subsequent inhibition of lipid synthesis and tumor progression (Zhang et al., 2016a). Future evaluation of the lifespan- and healthspan-promoting effects of SB-204990 must determine its potential as a

CRM. The ACLY and HSP90 inhibitor radicicol (first isolated from the fungus *Monosporium bonorden*) exhibits diverse protective effects in rodents, e.g., against renal and myocardial ischemia-reperfusion damage, but its pro-autophagic potential remains elusive (Sonoda et al., 2010).

Further synthetic agents capable of depleting AcCoA have been proposed as CRMs. Perhexiline maleate reduces AcCoA levels via inhibition of carnitine O-palmitoyl transferase 1 and is able to reversibly inhibit mTORC1 signaling and to promote autophagy *in vitro* (Balgı et al., 2009). It is a clinically approved anti-anginal agent that exhibits cardioprotective (Phan et al., 2009) and anti-tumor potential (Vella et al., 2015), although putative hepato- and/or neurotoxic effects need to be explored. Furthermore, UK5099 (a mitochondrial-pyruvate-carrier inhibitor) and 1,2,3-benzenetricarboxylate (an inhibitor of citrate transport) cause AcCoA depletion, protein deacetylation, and autophagy (Mariño et al., 2014). However, their *in vivo* effects need further investigation. Similarly, the impact of the pyruvate dehydrogenase inhibitor CPI-613 (a synthetic lipoate analog) on autophagy requires further investigation. CPI-613 has been shown to be tolerable in humans and to exhibit potential anti-tumor and chemotherapy-potentiating activity at pre-clinical and clinical levels (Alistar et al., 2017), whereas a recent phase II trial on small cell lung carcinoma patients failed to show beneficial effects.

NAD⁺ Intermediates

NAD⁺ concentrations decrease with age in rodents and humans at the systemic level, correlating with the development of age-associated pathologies (Das et al., 2018). Importantly, the overexpression of the NAD⁺-generating enzymes CYB5R3 and NQO1 is sufficient to increase murine life- and healthspan (Diaz-Ruiz et al., 2018). Interestingly, supplementation of NAD⁺ precursors, in particular nicotinamide riboside (NR) and nicotinamide mononucleotide (NMN), also exerts anti-aging effects (Das et al., 2018). For instance, in rodents, NMN and NR have been shown to cause hepato- and cardioprotective effects (Rajman et al., 2018); alleviate vascular aging (Das et al., 2018); improve learning, memory, and cognitive function in Alzheimer disease models; promote muscle function in models of muscular dystrophy (Rajman et al., 2018); and ameliorate diabetic pathophysiology (Yoshino et al., 2011), among other effects. In fact, NR could extend the lifespan of mice even when administered late in life (Zhang et al., 2016c). NMN and NR are contained in a variety of daily natural food sources, including different vegetables, fruits, meat, and shrimp as well as in human milk.

Nicotinamide (NAM, also called vitamin B3), another NAD⁺ precursor, can prevent aging-associated glaucoma in a mouse model (Williams et al., 2017). Chronic feeding with NAM on a low-fat diet or HFD fails to improve longevity but promotes the healthspan of aged mice (Mitchell et al., 2018). Specifically, glucose homeostasis, body fat percentage (on a low-fat diet), and locomotor activity (on HFD) were improved, while steatosis and inflammation were reduced (on HFD) (Mitchell et al., 2018).

Besides direct supplementation of NAD⁺ metabolites and precursors, beneficial elevation of NAD⁺ levels may also be achieved by interacting with its intracellular generation. A recent report shows that the pharmacological inhibition of the enzyme α -amino- β -carboxymuconate- ϵ -semialdehyde decarboxylase (ACMSD), which is a limiting step in *de novo* NAD⁺ synthesis in

the kidney and the liver, might be another possible mode of action for potential CRM-like agents (Katsyuba et al., 2018). The authors also identified two specific inhibitors that could be delivered via chow supplementation (Katsyuba et al., 2018).

At the molecular level, NAD⁺ is required for SIRT1 deacetylase activity and is thus instrumental for protein-deacetylation-mediated autophagy induction in general. The NAD⁺/SIRT1 route reportedly upregulates autophagy via deacetylation of Atg5, Atg7, and Atg8 in murine neurons (Fang et al., 2017) and stimulates mitophagy (Fang et al., 2017). In addition, NAD⁺ has been suggested to induce autophagy via AMPK (Fang et al., 2017). Preclinical safety assessments in rodents have suggested no adverse response upon short- or long-term treatment with NR or NMN (Fang et al., 2017). In the first controlled clinical trial of NR, good bioavailability and increased blood levels of NAD⁺ could be detected (Trammell et al., 2016). NAM has been shown in a large phase III trial to reduce the incidence of non-melanoma skin cancers (Chen et al., 2015). Interestingly, treatment with the nicotinic acid analog acipimox (a medication against hyperlipidemia) improved skeletal muscle mitochondrial function in type 2 diabetes patients (van de Weijer et al., 2015). A number of clinical trials are currently underway to assess the efficacy of NAD⁺ precursors in humans, especially of NR.

Aspirin

The non-steroidal anti-inflammatory drug acetylsalicylic acid, better known as aspirin, has been in extensive medical use since 1899. Before that, salicylates from willow bark were widely used in folk medicine. Aspirin, which quickly metabolizes to salicylate *in vivo*, has lifespan-increasing effects on model organisms, including mice (Strong et al., 2008), but seems to inhibit growth in yeast (Baroni et al., 2018; Carmona-Gutierrez et al., 2018; Madeo et al., 1997).

Salicylate inhibits EP300 by competing with acetyl-CoA, thus activating autophagy (Pietrocola et al., 2018). Aspirin also reduces mTOR signaling and activates AMPK and autophagy in colorectal cancer (CRC) cells (Din et al., 2012), which might explain its anti-cancer efficacy. However, contradictory results were reported on cardiac fibroblasts, in which it inhibited autophagy (Liu et al., 2017b). AMPK activation was also found *in vivo* in mice (Hawley et al., 2012) and might represent a major mechanism of aspirin-triggered CRM effects. Recently, aspirin has been suggested as an anti-cancer therapeutic (Patrignani and Patrono, 2016), as continuous intake is associated with lower tumor occurrence and decreased metastasis of CRC and breast and prostate cancer in humans (Patrignani and Patrono, 2016). Mortality-reducing effects were found in several human studies after the long-term use of low doses (3–10 years; circa 75–300 mg/day). Several meta-analyses have suggested aspirin as a primary and secondary prevention therapy of CVD, reducing both the risk for CVDs and mortality. In line, aspirin provided heart protection and improved glucose tolerance in rodents (Liu et al., 2017b). Additionally, mitophagy was induced in cardiomyocytes (Pietrocola et al., 2018) and fat-induced insulin resistance improved in mice (Kim et al., 2001), though opposite effects in humans have been reported (Netea et al., 2001).

The positive effects of long-term aspirin intake might outweigh the risk of negative ones, e.g., gastrointestinal bleeding, if it became possible to exclude patients at risk. Overall, aspirin seems a reasonable and cost-efficient CRM with a high thera-

peutic potential against multiple diseases but a generally low risk for human health.

(Poly)phenols

In general, phenolic compounds may represent an attractive source of (potential) CRMs. Epidemiological studies have linked an elevated intake of polyphenol-rich food (mostly fruits and vegetables) and drinks (including coffee, tea, and wine) to a reduced incidence of malignant, cardiovascular, and neurodegenerative diseases (Vauzour et al., 2010). Caffeic acid (CA; found in the eucalyptus bark) and gallic acid (GA; found in black tea and many plants), including derivatives thereof, for instance, reportedly induce autophagy (Doan et al., 2015), AMPK activation (Doan et al., 2015; Tyszkka-Czochara et al., 2017), protein deacetylation (Pietrocola et al., 2012), extended longevity across species, and anti-diabetes effects (Eid et al., 2017). While GA was found to inhibit EP300 (Lee et al., 2015c), CA might activate SIRT1s, namely SIRT3 (Mu et al., 2015). Moreover, CA has been shown to induce autophagy and improve glucose and lipid metabolism as well as renal function in a diabetic rat model (Matboli et al., 2017). Notably, CA phenyl ester (CAPE), an AMPK activator, has broad health-promoting properties and is a major bioactive component of propolis, a honeybee product commonly used in traditional medicine. CAPE has been reported to extend the lifespan of a mouse model of amyotrophic lateral sclerosis (ALS) (Fontanilla et al., 2012). Additionally, an ethanamide derivative (CAEA) has been shown to activate AMPK as well as SIRT1s and ameliorate cardiac damage in a mouse model (Lee et al., 2015b).

Another example is the stilbenoid piceatannol (found, e.g., in passion fruit seeds), an analog of resveratrol, which—along with its metabolite isorhapontigenin—was shown to stimulate SIRT1, activate autophagy (synergistically with resveratrol), deacetylate cytosolic proteins, improve parameters of metabolic syndrome in obese mice, promote murine astrocyte differentiation *in vivo*, and extend the lifespan of worms (Pietrocola et al., 2012; Surh and Na, 2016). However, to our knowledge, lifespan and healthspan data of piceatannol in higher models are elusive.

Curcumin is the major polyphenol in the rhizome of turmeric (*Curcuma longa*) and has a long tradition as a medical herb. Indeed, curcumin feeding extends the lifespan of non-rodent models and exerts cardioprotective, antineoplastic, and antidiabetic effects in rodent models (Rahmani et al., 2018). Dietary curcumin is readily metabolized to tetrahydrocurcumin (THC), which was shown to prolong the lifespan of middle-aged mice (Kitani et al., 2007). Numerous clinical trials have aimed at assessing the health effects of curcumin on humans, with positive effects reported for multiple diseases including different types of cancer, metabolic syndrome, depression, and diabetes (Zheng et al., 2018). Curcumin seems well tolerated and non-toxic. Its poor bioavailability can be significantly increased by several agents, including piperine (Shoba et al., 1998) (a major component in black pepper). The mode of action of curcumin remains to be clarified and—as with other polyphenols—may involve antioxidant properties but also autophagy induction, at least in some pathological settings. Its pro-autophagic activity has been connected to AMPK activation (Xiao et al., 2013) and mTOR signaling (Wang et al., 2014) as well as to inhibition of EP300 (Pietrocola et al., 2015).

A number of polyphenols are known to act as EP300 inhibitors, associated with autophagy induction and health-promoting

effects. These include anacardic acid (AC) (Pietrocola et al., 2015) (from the nutshell of the cashew, *Anacardium occidentale*) and garcinol (Pietrocola et al., 2015) (from the fruit of the Kokum tree, *Garcinia indica*). Similarly, the synthetic EP300 inhibitor C646 induces autophagy (Pietrocola et al., 2015) and exerts protective effects, including immunostimulatory antitumor activity (Liu et al., 2013). However, histone acetyl transferase (HAT) selectivity might be compromised at higher concentrations (van den Bosch et al., 2016). Interestingly, a novel EP300 inhibitor (A-485) shows higher potency as well as HAT selectivity and may suppress tumor growth (Lasko et al., 2017).

Several flavonoids, a multifunctional and highly bioactive polyphenolic subclass, which comprises more than 5,000 plant-derived substances, also promote autophagy coupled to protein deacetylation (Pietrocola et al., 2012). For instance, quercetin (*inter alia* found in black tea, onions, rocket, cole, curcuma, and fruits) and the nutritionally less abundant myricetin (sources: black tea, cole, parsley, garlic, curcuma, and fruits), which only differ in the position of a hydroxy group, were shown to induce autophagy and reduce protein acetylation to the same extent (Pietrocola et al., 2012). Both agents activate SIRT1 (D'Andrea, 2015; Jung et al., 2017) and extend lifespan in worms (DAF-16 dependent) (Büchter et al., 2015; Pietsch et al., 2009). They also promote the survival of neurodegenerative fly models (Ara et al., 2017; Kong et al., 2016), while only quercetin was shown to increase the lifespan of wild-type *Drosophila* (Proshkina et al., 2016). Quercetin supplementation in mice did not cause beneficial effects on longevity (though this was only tested in combination with polyphenolic taxifolin and pycnogenol) (Spindler et al., 2013). However, in combination with the chemotherapeutic dasatinib, quercetin acted senolytically, thus removing senescent cells *in vivo*, and improved cardiac function of aged mice while overall promoting the healthspan (Zhu et al., 2015). Of note, genetic ablation of senescent cells by means of an inducible suicide gene can extend rodent lifespan up to 25% (Baker et al., 2016). However, the contribution of senescent cells to aging progression—although well explored—remains to be fully understood and incorporated into an applicable model (van Deursen, 2014). In fact, the concept of senolytic drugs has only recently been developed, and more extensive mechanistic studies are needed to strengthen and elaborate the idea. For instance, the above-mentioned studies suggesting quercetin (in combination with other agents) to be senolytic but to fail in extending lifespan (Spindler et al., 2013; Zhu et al., 2015) may reflect a greater contribution of senolytic activity to healthspan than to lifespan-extension. However, differences in the agents used for combinations, application, dosage, timing, and mouse strains among other factors in these two studies underline that further analyses are warranted to evaluate possible pro-longevity effects of quercetin.

Furthermore, *in vivo* anti-cancer and anti-inflammatory effects as well as improvements of insulin sensitivity and HFD-induced weight gain have been described for quercetin (Chen et al., 2016; D'Andrea, 2015). Only slow progress has been made in translating these results to clinical trials: ambiguous results of quercetin supplementation on inflammation were reported. Quercetin improved blood pressure in hypertension, obesity, and type 2 diabetes patients (D'Andrea, 2015), also reducing plasma levels of oxidized low-density lipoprotein (Egert et al.,

2009). Although safe for human application, quercetin bioavailability is low and metabolization is high (D'Andrea, 2015), two features that might reduce its clinical utility.

The flavonoids (+)-catechin and its cis-form (–)-epicatechin can be prominently found in cocoa, tea brews, and red wine. Several animal studies and human trials have shown cardiovascular protective properties of catechins (Aprotosoaie et al., 2016). Both catechins reduce cytosolic protein acetylation and induce autophagy (Pietrocola et al., 2012), likely via pleiotropic targets. Epicatechin was suggested to act in a SIRT1-independent fashion (de Boer et al., 2006), and its dietary supplementation in wild-type flies and obese diabetic mice reduced mortality in both species and improved markers of systemic inflammation and diabetes-associated liver and aorta degeneration in the latter (Si et al., 2011). This is of special interest since epicatechin has been suggested as an insulin receptor activator by *in silico* analyses (Ganugapati et al., 2011). Catechin also improves stress resistance and extends the lifespan of nematodes, independently of antioxidative properties (Saul et al., 2011). The bioavailability of epicatechin appears favorable (Steffen et al., 2008) and numerous clinical trials have proven basic safety in patients. Epicatechin was shown to improve insulin resistance (Dower et al., 2015) and alter gene expression profiles, slightly downregulating inflammation- and adipogenesis-associated genes (Esser et al., 2018). Interestingly, epicatechin supplementation improved CVD markers and reduced triglyceride levels in patients with hypertriglyceridemia (Gutiérrez-Salmeán et al., 2016). However, many studies use chemically non-defined catechin-enriched (tea) extracts, rendering their evaluation problematic.

Epigallocatechin-3-gallate (EGCG) is the major polyphenol in green tea. It has numerous biological effects and pleiotropic molecular targets. Notably, it strongly inhibits HATs, activates SIRT1 (Lee et al., 2015a), and extends lifespan in worms (Abbas and Wink, 2009), flies (Wagner et al., 2015), and rats (Niu et al., 2013). It induces autophagy in cell culture in a reportedly Ca^{2+} /calmodulin-dependent protein kinase kinase beta (CaMKK β)-dependent fashion (Kim et al., 2013) and inhibits EP300/CBP (Ko et al., 2013). However, the exact role of protein deacetylation in EGCG-mediated pro-autophagic effects remains unclear. Additionally, reduced glucose metabolism and increased fitness of flies (Wagner et al., 2015) were reported. In rodents, EGCG improved liver and kidney function and reduced NF- κ B signaling (Niu et al., 2013). EGCG may stimulate AMPK, probably via the activation of the upstream kinase Ca^{2+} /calmodulin-dependent protein kinase kinase (CaMKK) (Collins et al., 2007). Several studies found promising effects of EGCG on the reduction of obesity and high-calorie-associated effects in rodents (Legeay et al., 2015). A 2013 study, however, did not find changes in metabolism, body weight, or liver function in nutritionally supplemented obese women (Mielgo-Ayuso et al., 2014). EGCG has also been suggested as a potential anti-cancer therapeutic (Du et al., 2012) and improves insulin sensitivity, glucose metabolism, and endothelial function in mice (Legeay et al., 2015). Prominently, EGCG was intensively studied in the context of neurodegeneration (including in clinical trials), as its anti-aggregation properties hold great promise (Cascella et al., 2017). Of note, a green tea extract (31.7 % EGCG, 8.5 % epicatechin) was neuroprotective and improved learning capacities in a

progeroid mouse model when fed lifelong or from adulthood (Unno et al., 2009). In humans, EGCG bioavailability is rather poor, challenging the utility of its clinical use (Legeay et al., 2015). Moreover, it has been reported that continuous intake of more than 800 mg per day may cause liver toxicity in patients (Hu et al., 2018). Finally, the alkaloid polyphenol berberine activates autophagy via SIRT1 and reduces protein acetylation (Shukla et al., 2016), protecting liver function (Sun et al., 2018), exerting neuroprotective effects (Wang et al., 2017), and reducing cardiac damage (Yu et al., 2016) in rodents. Berberine was also shown to extend the lifespan of *Drosophila* (Navrotskaya et al., 2012). Though oral bioavailability of berberine is poor (Liu et al., 2016), several clinical studies have been or are being performed with berberine supplementation. Noteworthy, berberine showed hypoglycemic effects and improved insulin parameters in type 2 diabetes patients (Yin et al., 2008).

In general, polyphenols represent a chemical group with a great potential to find pharmacological alternatives to CR. For instance, a recent study screening for pro-longevity drugs identified the flavonoid 4,4'-dimethoxychalcone as a pro-autophagic natural compound (present in the plant *Angelica keiskei*, also known as Ashitaba). 4,4'-Dimethoxychalcone can extend lifespan from yeast to flies and shows cardioprotective effects in mice, all in an autophagy-dependent manner (Carmona-Gutierrez et al., 2019).

It is important to note that most polyphenols possess antioxidant properties and have pleiotropic effects on several molecular targets, rendering it difficult to study their precise mode of action. Moreover, modifications such as glycosylation are likely to change their bioactive features *in vivo*. For instance, glycosylation greatly modulates the effects of quercetin on the worm lifespan and probably alters its bioavailability (Pallauf et al., 2017).

Metformin

Metformin (dimethylbiguanide hydrochloride) is a derivative of natural guanidines present in the French lilac (*Galega officinalis*), a plant that has been used in folk medicine for centuries. Originally described as a hypoglycemic and antimalarial drug, it is currently a widely prescribed agent in the treatment of type 2 diabetes. Interestingly, metformin administration extends the lifespan in different animal models, including mammals (Martin-Montalvo et al., 2013). In humans, metformin seems to be beneficial against a number of age-related diseases, including cancer and metabolic syndrome as well as cognitive and cardiovascular disorders (Foretz et al., 2014; Greenhill, 2015). Indeed, a recent meta-analysis of diabetics on metformin use has revealed that this drug reduces all-cause mortality and age-associated diseases (Campbell et al., 2017). This geroprotective potential paired with its little side effects has propelled the start of several clinical trials. Metformin recapitulates important metabolic effects of CR (Onken and Driscoll, 2010) and stimulates protective autophagy, for instance in mouse models of obesity and cardiac dysfunction (Li et al., 2017a; Xie et al., 2011).

Mechanistically, metformin has been associated with the activation of the master energy sensor AMPK (Duca et al., 2015) via the inhibition of the mitochondrial electron transport chain complex I (Owen et al., 2000), although it might activate AMPK through a lysosomal pathway, as well (Zhang et al., 2016b). In addition, metformin also inhibits mTORC1 indepen-

dently of AMPK (Nair et al., 2014). Whether metformin effects rely on protein hypoacetylation remains to be systematically evaluated. In fact, metformin-mediated AMPK activation has been associated with both reduced EP300 and CREB-binding protein (He et al., 2009; Lim et al., 2012) and increased HAT (HAT1) activity (Marin et al., 2017). Similarly, metformin can inhibit class II HDACs (Khan and Jena, 2016) but can also stimulate class III HDAC SIRT1 activity, possibly downstream of AMPK activation (Caton et al., 2010). Metformin may also impact SIRT1 gene expression directly. Altogether the current data suggest that metformin could qualify as a *bona fide* CRM if causal effects on protein hypoacetylation were validated.

Rapamycin and Related Compounds (Rapalogs)

Rapamycin (sirolimus) is a macrolide compound produced by *Streptomyces hygroscopicus*. It was originally used as an antifungal drug and is an FDA-approved immunosuppressant that has been shown to extend lifespan in *C. elegans*, *D. Melanogaster*, and mice (Ehninger et al., 2014). Rapamycin has also been connected, for example, to cardioprotection (Chiao et al., 2016), anti-neurodegenerative effects (Kolosova et al., 2013), and obesity prevention (Chang et al., 2009) in rodents. Numerous clinical trials have addressed the efficacy of rapamycin and rapamycin analogs (rapalogs) in treating diseases, including cancers. Rapamycin can inhibit mTORC1 by forming a complex with the protein FKBP12 (Boutouja et al., 2019). As a specific mTORC1 inhibitor, rapamycin promotes autophagy independently of SIRT1 (Kim and Guan, 2015) but links to possible autophagy-relevant deacetylation processes have been documented (Füllgrabe et al., 2013). Whether such deacetylation processes contribute to the beneficial effects of rapamycin will clarify if it fully qualifies as a CRM. However, its immunosuppressant properties compromise the broad clinical application. In addition, prolonged rapamycin treatment exacerbates insulin resistance and diabetes (Lamming et al., 2012) and actually reduces the lifespan of diabetic mice (Sataranatarajan et al., 2016). Whether intermittent administration of rapamycin might circumvent these adverse effects needs further investigation.

Rapamycin belongs to the so-called first generation mTOR inhibitors, which also comprise the rapalogs temsirolimus and everolimus, both of which also bind FKBP12 but show improved pharmacokinetics (Boutouja et al., 2019). While the first generation rapalogs and rapamycin only block mTORC1, the second generation of mTOR inhibitors (NVP-BE235, PF-04691502, OSI-027, and others) acts by blocking the ATP site of the mTOR kinase, thus also affecting TORC2 (Boutouja et al., 2019). A series of clinical trials for specific medical applications have been conducted or registered for some of these drugs, but further evaluation is required to assess whether they also possess health- and/or lifespan-extending features. Finally, the third generation of mTOR inhibitors includes bivalent drugs that target multiple molecular targets in the TOR complexes (e.g., mTOR kinase, FRB domain, and FKBP12), providing enhanced effectivity against, for instance, tumorous cells (Boutouja et al., 2019; Fan et al., 2017). One recent example of this category is Rapalink-1, which is a specific TOR kinase inhibitor linked to rapamycin (Fan et al., 2017). This generation, however, has not surfaced to clinical studies yet.

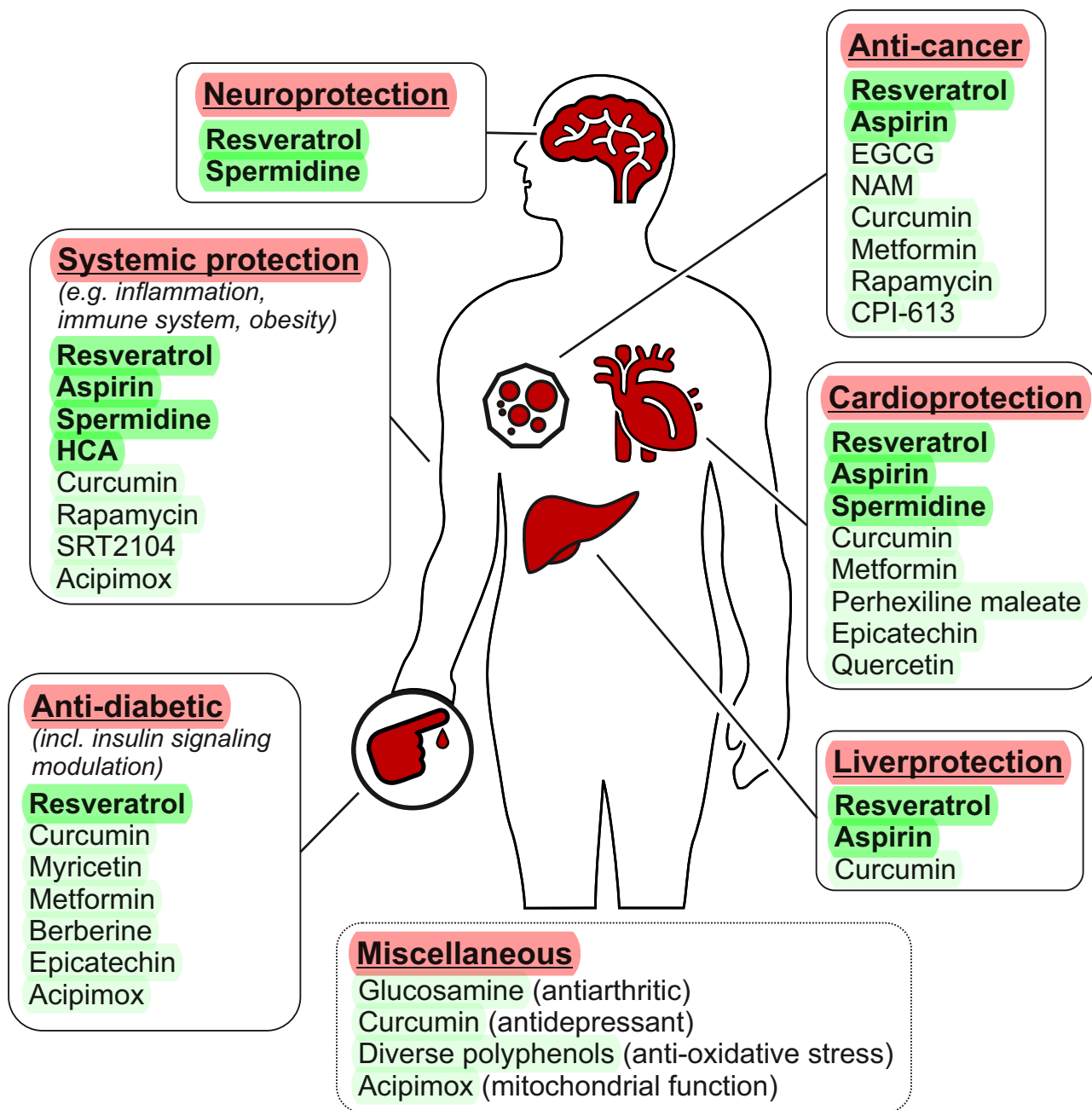


Figure 3. Possible Physiological Effects of CRMs Seen in Clinical Studies

CRMs positively affect human health both at the systemic and tissue-specific levels. Bold, CRMs; normal, potential CRMs. EGCG, Epigallocatechin-3-gallate; HCA, hydroxycitric acid; NMN, nicotinamide mononucleotide; NAM, nicotinamide.

Modulation of Glucose Metabolism

Blocking cellular energy utilization, specifically glycolysis, has been devised as a CRM strategy. Glucose deprivation alone is sufficient to induce autophagy via AMPK/mTOR (Moruno et al., 2012). The hexokinase inhibitor glucosamine is a widely used agent to prevent and treat osteoarthritis and shows no relevant side effects. In fact, glucosamine medication has been associated with decreased mortality in humans (Bell et al., 2012). Autophagy activation by glucosamine was proposed to be

both mTOR dependent (Caramés et al., 2013) and independent (Shintani et al., 2010), meaning that the exact pathway is still debated. Whether protein deacetylation is involved in glucosamine-mediated autophagy induction remains to be studied. Instead, 2-deoxyglucose (a synthetic hexokinase 2 inhibitor) increases nematode lifespan but seems to be cardiotoxic and to increase mortality in rats (Minor et al., 2010). In line with the latter results, 2-deoxyglucose treatment suppresses autophagy via activation of mTORC1 (Roberts et al., 2014) and does not

Table 2. Effects of CRMs, Potential CRMs, and Other Compounds on Rodent Lifespan

Group	Substance	Organism	Lifespan Extension	Application Scheme	References
CRMs	aspirin (and salicylate)	mouse (male UM-HET3; failed in females)	~8% median lifespan increase	starting at 4 months; supplemented via Purina 5LG6 diet	Strong et al., 2008
	resveratrol	mouse (male C57BL/6NIA)	31% reduced risk of death	starting at 12 months; supplemented via diet; on high-calorie diet	Baur et al., 2006
	spermidine	mouse (male and female C57BL/6J)	~12% or ~10% median lifespan increases for lifelong or late-in-life supplementation, respectively	starting at 4 or 18 months; supplemented via drinking water; standard diet	Eisenberg et al., 2016
Potential CRMs	curcumin (or tetrahydrocurcumin)	mouse (male C57BL/6)	11.7% mean lifespan increase	starting at 13 months; supplemented via standard diet	Kitani et al., 2007
	epicatechin	mouse (db/db obese, diabetic model)	reduced mortality (8.4% in treated versus 50% in control group after 15 weeks of treatment)	starting at 5 weeks; supplemented via drinking water	Si et al., 2011
	EGCG	rat (male Wistar)	~13.5% median lifespan increase	starting at 5 weeks; supplemented via drinking water	Niu et al., 2013
	metformin	mouse (male C57BL/6 and B6C3F1)	5.83% (C57BL/6) and 4.15% (B6C3F1) mean lifespan increase for low dose (0.1%); high dose (1%) led to 14.4% mean lifespan reduction	starting at 54 weeks; supplemented via standard diet	Martin-Montalvo et al., 2013
	NR	mouse (C57BL/6J)	~5% mean lifespan increase	starting at 22–24 months; via diet; for 6 weeks; standard diet	Zhang et al., 2016c
	rapamycin	mouse (male and female C57BL/6, UM-HET3, 129/sv)	median (~25%) and maximum lifespan increase (Miller et al., 2014); 10% median lifespan increase (Anisimov et al., 2011); median lifespan increase of 10% (males) and 18% (females) (Miller et al., 2011); mean lifespan increase of 9% (male) and 13% (female) (Harrison et al., 2009)	many different application methods and starting timepoints of intervention reported	reviewed in Ehninger et al., 2014
	SRT1720	mouse (male C57BL/6J)	11% and 44% mean lifespan increase for low and high doses	starting at 12 months; supplemented via diet; on high-fat diet	Minor et al., 2011
Others	CAPE	mouse (male and female SOD1 ^{G93A} ALS model with B6SJL background)	~7% lifespan increase	single daily oral dose after disease onset	Fontanilla et al., 2012
	SRT2104	mouse (male C57BL/6J)	9.7% mean lifespan increase	starting at 6 months; supplemented via diet; on standard diet	Mercken et al., 2014

Lifespan experiments that have been performed in rodents with compounds listed in [Table 1](#). Animal specificities (including sex), quantitation of lifespan improvement, experimental design for compound administration, and corresponding references are noted. CAPE, caffeic acid phenyl ester; EGCG, epigallocatechin-3-gallate; NR, nicotinamide riboside.

represent a CRM. Acarbose is an α -glucosidase inhibitor of bacterial origin (*Streptomyces* and *Actinoplanes* species) and widely used as an anti-diabetic medication, preventing the release of glucose from more complex carbohydrates (Brewer et al., 2016). Although it might be connected to further lifespan-determining effects (Harrison et al., 2014), additional studies that address its possible influence on autophagy and protein deacetylation are needed.

Conclusion

Ongoing and future clinical trials, as well as meta-analyses, will ultimately determine the actual beneficial impact of each CRM on human health (Figure 3). Further CRMs may be discovered and optimized versions of known CRMs obtained by medicinal chemistry that will need to be further evaluated. Besides, it will be important to tackle certain limitations (e.g., bioavailability) and unfold possibilities, including the prospect for combinatorial approaches. Many CRMs fail to extend lifespan to the same degree as CR or fasting does and some CRMs show sex-specific differences (Table 2), suggesting that CR might cumulatively ignite distinct pathways that are only partly targeted by single CRMs. This propels the idea of achieving additive effects by compound/treatment combinations. This applies to (1) combinations of distinct CRMs, (2) combinations of CRMs with other beneficial non-CRM compounds, and (3) combinations of CRMs with behavioral/nutritional approaches (e.g., fasting, CR, and exercise). With respect to (1), it can be expected that CRMs that act on distinct routes to achieve protein deacetylation (namely AcCoA depletion, acetyltransferase inhibition, or deacetylase activation) could interact in a synergistic fashion. For example, resveratrol (which promotes SIRT deacetylase activity) synergizes with spermidine (which is an acetyltransferase inhibitor) to promote autophagy *in vitro* (human cell culture) and *in vivo* (mice) (Morselli et al., 2011). Moreover, rapamycin and metformin act synergistically on worm lifespan (Admasu et al., 2018). These studies exemplify the substantial potential of such combinatorial approaches in the anti-aging field. Regarding (2), several health-promoting compounds including antioxidant and hormesis mimetics do not rely on the deacetylation-autophagy axis. It will be interesting to investigate whether they might be favorably combined with CRMs. Finally, (3) exercise, CR, fasting, and CRMs all promote autophagy, and additive or synergistic effects might be attained upon combining these interventions. For instance, several studies suggest that exercise might be combined with the CRM resveratrol (Liao et al., 2017).

Another important aspect is the timing of CRM application. First, the effective administration of CRMs in a middle-life stage (instead of lifelong application), before adverse age-associated symptoms manifest, would greatly enhance the therapeutic feasibility of CRMs for humans. Thus, exploring whether specific CRMs can be effective also upon administration late in life will help clarify the extended potential of these drugs. Second, the application of CRMs in a rhythmic fashion could reduce the drug load while promoting the same effects. This follows the idea that in humans, rhythmic variations of calorie intake (e.g., intermittent fasting) stimulate many of the favorable effects that constant CR does (Di Francesco et al., 2018). In fact, in animal and human studies, the timing of meals regarding the circadian clock is of utmost importance (Di Francesco et al., 2018). Third,

the target availability for different CRMs might be optimized by timing the administration of a given CRM with the expression patterns of its corresponding target(s). In fact, if a specific cellular target is not expressed at relevant levels, a drug dose at this time point might be ineffective. Mechanistic data on cellular targets of specific CRMs are already available and solid. The experiments exploring “target expression-enhanced” administration might be especially interesting regarding variable expression profiles in different tissue types as well as with ongoing age and at a particular disease status.

Irrespective of the pending therapeutic validation of CRMs as a stand-alone and/or combinatory approach and other open questions, the available data substantiate the large potential of pharmacological autophagy induction as a feasible and effective strategy against multiple diseases.

ACKNOWLEDGMENTS

F.M. is grateful to the Austrian Science Fund FWF (Austria) for grants P23490-B20, P29262, P24381, P29203 P27893, I1000, and “SFB Lipotox” (F3012), as well as to Bundesministerium für Wissenschaft, Forschung und Wirtschaft, and the Karl-Franzens University for grant “Unkonventionelle Forschung” and grant DKplus Metabolic and Cardiovascular Diseases (W1226). We acknowledge support from NAWI Graz and the BioTechMed-Graz flagship project “EPIAge.” G.K. is supported by the Ligue contre le Cancer Comité de Charente-Maritime (équipe labellisée); Agence National de la Recherche (ANR)–Projets blancs; ANR under the frame of E-Rare-2, the ERA-Net for Research on Rare Diseases; Association pour la recherche sur le cancer (ARC); Cancéropôle Ile-de-France; Institut National du Cancer (INCa); Institut Universitaire de France; Fondation pour la Recherche Médicale (FRM); the European Commission (ArtForce); the European Research Council (ERC); the LeDucq Foundation; the LabEx Immuno-Oncology; the Recherche Hospitalo-Universitaire Torino Lumière, the Site de Recherche Intégrée sur le Cancer (SIRIC) Stratified Oncology Cell DNA Repair and Tumor Immune Elimination (SOCRATE); the SIRIC Cancer Research and Personalized Medicine (CARPEM); and the Paris Alliance of Cancer Research Institutes (PACRI).

AUTHOR CONTRIBUTIONS

F.M., D.C.-G., S.J.H., and G.K. contributed to the conceptualization, figure design, and writing of the manuscript.

DECLARATION OF INTERESTS

G.K. is inventor on a patent application (WO2015049365 A3) submitted by INSERM (Institut National de la Santé et de la Recherche Médicale), Assistance Publique-Hôpitaux De Paris (APHP), Université Paris Descartes, Université Pierre et Marie Curie (Paris 6), Université Paris Diderot-Paris 7, Université Paris-Sud, Institut Gustave Roussy, that covers the medical use of CRMs. F.M. and D.C.-G. have equity interests in TLL (The Longevity Labs), a company founded in 2016 that will develop natural food extracts. F.M., D.C.-G., and G.K. are the scientific co-founders of Samsara Therapeutics.

REFERENCES

- Abbas, S., and Wink, M. (2009). Epigallocatechin gallate from green tea (*Camellia sinensis*) increases lifespan and stress resistance in *Caenorhabditis elegans*. *Planta Med* 75, 216–221.
- Admasu, T.D., Batchu, K.C., Barardo, D., Ng, L.F., Lam, V.Y.M., Xiao, L., Cazenave-Gassiot, A., Wenk, M.R., Tolwinski, N.S., and Gruber, J. (2018). Drug synergy slows aging and improves healthspan through IGF and SREBP lipid signaling. *Dev. Cell* 47, 67–79.e5.
- Alistar, A., Morris, B.B., Desnoyer, R., Klepin, H.D., Hosseinzadeh, K., Clark, C., Cameron, A., Leyendecker, J., D’Agostino, R., Topaloglu, U., et al. (2017). Safety and tolerability of the first-in-class agent CPI-613 in combination with modified FOLFIRINOX in patients with metastatic pancreatic cancer: a single-centre, open-label, dose-escalation, phase 1 trial. *Lancet Oncol.* 18, 770–778.

Anisimov, V.N., Zabezhinski, M.A., Popovich, I.G., Piskunova, T.S., Semenchenko, A.V., Tyndyk, M.L., Yurova, M.N., Rosenfeld, S.V., and Blagosklonny, M.V. (2011). Rapamycin increases lifespan and inhibits spontaneous tumorigenesis in inbred female mice. *Cell Cycle* 10, 4230–4236.

Aprotosoaie, A.C., Miron, A., Trifan, A., Luca, V.S., and Costache, I.I. (2016). The cardiovascular effects of Cocoa polyphenols—an overview. *Diseases* 4, 39.

Ara, G., Afzal, M., Jyoti, S., and Siddique, Y.H. (2017). Effect of myricetin on the transgenic *Drosophila* model of Parkinson's disease. *Bull. Fac. Pharm. Cairo Univ.* 55, 259–262.

Ard, J.D., Gower, B., Hunter, G., Ritchie, C.S., Roth, D.L., Goss, A., Wingo, B.C., Bodner, E.V., Brown, C.J., Bryan, D., et al. (2017). Effects of calorie restriction in obese older adults: the CROSSROADS randomized controlled trial. *J. Gerontol. A Biol. Sci. Med. Sci.* 73, 73–80.

Asghar, M., Monjok, E., Kouamou, G., Ohia, S.E., Bagchi, D., and Lokhandwala, M.F. (2007). Super CitriMax (HCA-SX) attenuates increases in oxidative stress, inflammation, insulin resistance, and body weight in developing obese Zucker rats. *Mol. Cell. Biochem.* 304, 93–99.

Baker, D.J., Childs, B.G., Durik, M., Wijers, M.E., Sieben, C.J., Zhong, J., Saltness, R.A., Jeganathan, K.B., Verzosa, G.C., Pezeshki, A., et al. (2016). Naturally occurring p16^{INK4a}-positive cells shorten healthy lifespan. *Nature* 530, 184–189.

Balgi, A.D., Fonseca, B.D., Donohue, E., Tsang, T.C.F., Lajoie, P., Proud, C.G., Nabi, I.R., and Roberge, M. (2009). Screen for chemical modulators of autophagy reveals novel therapeutic inhibitors of mTORC1 signaling. *PLoS One* 4, e7124.

Baroni, M.D., Colombo, S., and Martegani, E. (2018). Antagonism between salicylate and the cAMP signal controls yeast cell survival and growth recovery from quiescence. *Microb. Cell* 5, 344–356.

Bauer, J.H., Morris, S.N.S., Chang, C., Flatt, T., Wood, J.G., and Helfand, S.L. (2009). dSir2 and Dmp53 interact to mediate aspects of CR-dependent lifespan extension in *D. melanogaster*. *Aging* 1, 38–48.

Baur, J.A., Pearson, K.J., Price, N.L., Jamieson, H.A., Lerin, C., Kalra, A., Prabhu, V.V., Allard, J.S., Lopez-Lluch, G., Lewis, K., et al. (2006). Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 444, 337–342.

Bell, G.A., Kantor, E.D., Lampe, J.W., Shen, D.D., and White, E. (2012). Use of glucosamine and chondroitin in relation to mortality. *Eur. J. Epidemiol.* 27, 593–603.

Berman, A.Y., Motechin, R.A., Wiesenfeld, M.Y., and Holz, M.K. (2017). The therapeutic potential of resveratrol: a review of clinical trials. *NPJ Precis. Oncol.* 1, 35.

Biel, T.G., Lee, S., Flores-Toro, J.A., Dean, J.W., Go, K.L., Lee, M.H., Law, B.K., Law, M.E., Dunn, W.A., Zendejas, I., et al. (2016). Sirtuin 1 suppresses mitochondrial dysfunction of ischemic mouse livers in a mitofusin 2-dependent manner. *Cell Death Differ.* 23, 279–290.

Boutouja, F., Stiehm, C.M., and Platt, H.W. (2019). mTOR: a cellular regulator interface in health and disease. *Cell* 8, 18.

Brewer, R.A., Gibbs, V.K., and Smith, D.L. (2016). Targeting glucose metabolism for healthy aging. *Nutr. Healthy Aging* 4, 31–46.

Büchter, C., Ackermann, D., Honnen, S., Arnold, N., Havermann, S., Koch, K., and Wätjen, W. (2015). Methylated derivatives of myricetin enhance life span in *Caenorhabditis elegans* dependent on the transcription factor DAF-16. *Food Funct.* 6, 3383–3392.

Burnett, C., Valentini, S., Cabreiro, F., Goss, M., Somogyvári, M., Piper, M.D., Hoddinott, M., Sutphin, G.L., Leko, V., McElwee, J.J., et al. (2011). Absence of effects of Sir2 overexpression on lifespan in *C. elegans* and *Drosophila*. *Nature* 477, 482–485.

Campbell, J.M., Bellman, S.M., Stephenson, M.D., and Lisy, K. (2017). Metformin reduces all-cause mortality and diseases of ageing independent of its effect on diabetes control: a systematic review and meta-analysis. *Ageing Res. Rev.* 40, 31–44.

Cantó, C., and Auwerx, J. (2011). Calorie restriction: is AMPK as a key sensor and effector? *Physiology (Bethesda)* 26, 214–224.

Caramés, B., Kiosses, W.B., Akasaki, Y., Brinson, D.C., Eap, W., Koziol, J., and Lotz, M.K. (2013). Glucosamine activates autophagy in vitro and in vivo. *Arthritis Rheum.* 65, 1843–1852.

Carmona-Gutierrez, D., Bauer, M.A., Zimmermann, A., Aguilera, A., Austriaco, N., Ayscough, K., Balzan, R., Bar-Nun, S., Barrientos, A., Belenky, P., et al. (2018). Guidelines and recommendations on yeast cell death nomenclature. *Microb. Cell* 5, 4–31.

Carmona-Gutierrez, D., Zimmermann, A., Kainz, K., Pietroccola, F., Chen, G., Maglioni, S., Schiavi, A., Nah, J., Mertel, S., Beuschel, C., et al. (2019). The flavonoid 4,4'-dimethoxychalcone promotes autophagy-dependent longevity across species. *Nat. Commun.* 4. Published online February 19, 2019. <https://doi.org/10.1038/s41467-019-08555-w>.

Cascella, M., Bimonte, S., Muzio, M.R., Schiavone, V., and Cuomo, A. (2017). The efficacy of Epigallocatechin-3-gallate (green tea) in the treatment of Alzheimer's disease: an overview of pre-clinical studies and translational perspectives in clinical practice. *Infect. Agent. Cancer* 72.

Caton, P.W., Nayuni, N.K., Kieswich, J., Khan, N.Q., Yaqoob, M.M., and Corder, R. (2010). Metformin suppresses hepatic gluconeogenesis through induction of SIRT1 and GCN5. *J. Endocrinol.* 205, 97–106.

Chacko, S.M., Thambi, P.T., Kuttan, R., and Nishigaki, I. (2010). Beneficial effects of green tea: a literature review. *Chin. Med.* 5, 13.

Chang, G.R., Chiu, Y.S., Wu, Y.Y., Chen, W.Y., Liao, J.W., Chao, T.H., and Mao, F.C. (2009). Rapamycin protects against high fat diet-induced obesity in C57BL/6J mice. *J. Pharmacol. Sci.* 109, 496–503.

Chen, A.C., Martin, A.J., Choy, B., Fernández-Peñas, P., Dalziel, R.A., McKenzie, C.A., Scolyer, R.A., Dhillon, H.M., Vardy, J.L., Krickler, A., et al. (2015). A phase 3 randomized trial of nicotinamide for skin-cancer chemoprevention. *N. Engl. J. Med.* 373, 1618–1626.

Chen, S., Jiang, H., Wu, X., and Fang, J. (2016). Therapeutic effects of quercetin on inflammation, obesity, and type 2 diabetes. *Mediators Inflamm.* 2016, 9340637.

Chiao, Y.A., Kolwicz, S.C., Basisty, N., Gagnidze, A., Zhang, J., Gu, H., Djukovic, D., Beyer, R.P., Raftery, D., MacCoss, M., et al. (2016). Rapamycin transiently induces mitochondrial remodeling to reprogram energy metabolism in old hearts. *Aging* 8, 314–327.

Chong, C.R., Sallustio, B., and Horowitz, J.D. (2016). Drugs that affect cardiac metabolism: focus on perhexiline. *Cardiovasc. Drugs Ther.* 30, 399–405.

Collins, Q.F., Liu, H.Y., Pi, J., Liu, Z., Quon, M.J., and Cao, W. (2007). Epigallocatechin-3-gallate (EGCG), a green tea polyphenol, suppresses hepatic gluconeogenesis through 5'-AMP-activated protein kinase. *J. Biol. Chem.* 282, 30143–30149.

Côté, C.D., Rasmussen, B.A., Duca, F.A., Zadeh-Tahmasebi, M., Baur, J.A., Daljeet, M., Breen, D.M., Filippi, B.M., and Lam, T.K.T. (2015). Resveratrol activates duodenal Sirt1 to reverse insulin resistance in rats through a neuronal network. *Nat. Med.* 21, 498–505.

Cuzick, J., Thorat, M.A., Bosetti, C., Brown, P.H., Burn, J., Cook, N.R., Ford, L.G., Jacobs, E.J., Jankowski, J.A., La Vecchia, C., et al. (2015). Estimates of benefits and harms of prophylactic use of aspirin in the general population. *Ann. Oncol.* 26, 47–57.

D'Andrea, G. (2015). Quercetin: a flavonol with multifaceted therapeutic applications? *FitoTerapia* 106, 256–271.

Dang, W. (2014). The controversial world of sirtuins. *Drug Discov. Today Technol.* 12, e9–e17.

Das, A., Huang, G.X., Bonkowski, M.S., Longchamp, A., Li, C., Schultz, M.B., Kim, L.J., Osborne, B., Joshi, S., Lu, Y., et al. (2018). Impairment of an endothelial NAD⁺-H₂S signaling network is a reversible cause of vascular aging. *Cell* 173, 74–89.e20.

de Boer, V.C.J., de Goffau, M.C., Arts, I.C.W., Hollman, P.C.H., and Keijer, J. (2006). SIRT1 stimulation by polyphenols is affected by their stability and metabolism. *Mech. Ageing Dev.* 127, 618–627.

Di Francesco, A., Di Germanio, C., Bernier, M., and de Cabo, R. (2018). A time to fast. *Science* 362, 770–775.

Diaz-Ruiz, A., Lanasa, M., Garcia, J., Mora, H., Fan, F., Martin-Montalvo, A., Di Francesco, A., Calvo-Rubio, M., Salvador-Pascual, A., Aon, M.A., et al. (2018).

- Overexpression of CYB5R3 and NQO1, two NAD⁺-producing enzymes, mimics aspects of caloric restriction. *Aging Cell*, e12767.
- Din, F.V.N., Valanciute, A., Houde, V.P., Zibrova, D., Green, K.A., Sakamoto, K., Alessi, D.R., and Dunlop, M.G. (2012). Aspirin inhibits mTOR signaling, activates AMP-activated protein kinase, and induces autophagy in colorectal cancer cells. *Gastroenterology* *142*, 1504–1515.e3.
- Doan, K.V., Ko, C.M., Kinyua, A.W., Yang, D.J., Choi, Y.H., Oh, I.Y., Nguyen, N.M., Ko, A., Choi, J.W., Jeong, Y., et al. (2015). Gallic acid regulates body weight and glucose homeostasis Through AMPK activation. *Endocrinology* *156*, 157–168.
- Dower, J.I., Geleijnse, J.M., Gijbbers, L., Zock, P.L., Kromhout, D., and Hollman, P.C.H. (2015). Effects of the pure flavonoids epicatechin and quercetin on vascular function and cardiometabolic health: a randomized, double-blind, placebo-controlled, crossover trial. *Am. J. Clin. Nutr.* *101*, 914–921.
- Du, G.J., Zhang, Z., Wen, X.D., Yu, C., Calway, T., Yuan, C.S., and Wang, C.Z. (2012). Epigallocatechin gallate (EGCG) is the most effective cancer chemopreventive polyphenol in green tea. *Nutrients* *4*, 1679–1691.
- Dubé, J.J., Amati, F., Toledo, F.G.S., Stefanovic-Racic, M., Rossi, A., Coen, P., and Goodpaster, B.H. (2011). Effects of weight loss and exercise on insulin resistance, and intramyocellular triacylglycerol, diacylglycerol and ceramide. *Diabetologia* *54*, 1147–1156.
- Duca, F.A., Côté, C.D., Rasmussen, B.A., Zadeh-Tahmasebi, M., Rutter, G.A., Filippi, B.M., and Lam, T.K.T. (2015). Metformin activates a duodenal Ampk-dependent pathway to lower hepatic glucose production in rats. *Nat. Med.* *21*, 506–511.
- Egert, S., Bosity-Westphal, A., Seiberl, J., Kürbitz, C., Settler, U., Plachta-Danielzik, S., Wagner, A.E., Frank, J., Schrezenmeier, J., Rimbach, G., et al. (2009). Quercetin reduces systolic blood pressure and plasma oxidised low-density lipoprotein concentrations in overweight subjects with a high-cardiovascular disease risk phenotype: a double-blinded, placebo-controlled cross-over study. *Br. J. Nutr.* *102*, 1065–1074.
- Ehninger, D., Neff, F., and Xie, K. (2014). Longevity, aging and rapamycin. *Cell. Mol. Life Sci.* *71*, 4325–4346.
- Eid, H.M., Thong, F., Nachar, A., and Haddad, P.S. (2017). Caffeic acid methyl and ethyl esters exert potential antidiabetic effects on glucose and lipid metabolism in cultured murine insulin-sensitive cells through mechanisms implicating activation of AMPK. *Pharm. Biol.* *55*, 2026–2034.
- Eisenberg, T., Knauer, H., Schauer, A., Büttner, S., Ruckstuhl, C., Carmona-Gutierrez, D., Ring, J., Schroeder, S., Magnes, C., Antonacci, L., et al. (2009). Induction of autophagy by spermidine promotes longevity. *Nat. Cell Biol.* *11*, 1305–1314.
- Eisenberg, T., Abdellatif, M., Schroeder, S., Primessnig, U., Stekovic, S., Pendl, T., Harger, A., Schipke, J., Zimmermann, A., Schmidt, A., et al. (2016). Cardioprotection and lifespan extension by the natural polyamine spermidine. *Nat. Med.* *22*, 1428–1438.
- Eiyama, A., and Okamoto, K. (2015). PINK1/Parkin-mediated mitophagy in mammalian cells. *Curr. Opin. Cell Biol.* *33*, 95–101.
- Esser, D., Geleijnse, J.M., Matulalutapauw, J.C., Dower, J.I., Kromhout, D., Hollman, P.C.H., and Afman, L.A. (2018). Pure flavonoid epicatechin and whole genome gene expression profiles in circulating immune cells in adults with elevated blood pressure: A randomised double-blind, placebo-controlled, crossover trial. *PLoS One* *13*, e0194229.
- Fan, Q., Aksoy, O., Wong, R.A., Ilkhanizadeh, S., Novotny, C.J., Gustafson, W.C., Truong, A.Y.-Q., Cayanan, G., Simonds, E.F., Haas-Kogan, D., et al. (2017). A kinase inhibitor targeted to mTORC1 drives regression in glioblastoma. *Cancer Cell* *31*, 424–435.
- Fang, E.F., Lautrup, S., Hou, Y., Demarest, T.G., Croteau, D.L., Mattson, M.P., and Bohr, V.A. (2017). NAD⁺ in aging: molecular mechanisms and translational implications. *Trends Mol. Med.* *23*, 899–916.
- Fontana, L., Weiss, E.P., Villareal, D.T., Klein, S., and Holloszy, J.O. (2008). Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. *Aging Cell* *7*, 681–687.
- Fontanilla, C.V., Wei, X., Zhao, L., Johnstone, B., Pascuzzi, R.M., Farlow, M.R., and Du, Y. (2012). Caffeic acid phenethyl ester extends survival of a mouse model of amyotrophic lateral sclerosis. *Neuroscience* *205*, 185–193.
- Foretz, M., Guigas, B., Bertrand, L., Pollak, M., and Viollet, B. (2014). Metformin: From mechanisms of action to therapies. *Cell Metab.* *20*, 953–966.
- French, S.W. (2016). Chronic alcohol bingeing injures the liver and other organs by reducing NAD⁺ levels required for sirtuin's deacetylase activity. *Exp. Mol. Pathol.* *100*, 303–306.
- Füllgrabe, J., Lynch-Day, M.A., Heldring, N., Li, W., Struijk, R.B., Ma, Q., Hermanson, O., Rosenfeld, M.G., Klionsky, D.J., and Joseph, B. (2013). The histone H4 lysine 16 acetyltransferase hMOF regulates the outcome of autophagy. *Nature* *500*, 468–471.
- Ganugapati, J., Mukkavalli, S., and Sahithi, A. (2011). Docking studies of green tea flavonoids as insulin mimetics. *Int. J. Comput. Appl.* *30*, 5.
- Goudarzvand, M., Afraei, S., Yaslianifard, S., Ghiasy, S., Sadri, G., Kalvandi, M., Alinia, T., Mohebbi, A., Yazdani, R., Azarian, S.K., et al. (2016). Hydroxytyrosic acid ameliorates inflammation and oxidative stress in mouse models of multiple sclerosis. *Neural Regen. Res.* *11*, 1610–1616.
- Greenhill, C. (2015). Gastric cancer. Metformin improves survival and recurrence rate in patients with diabetes and gastric cancer. *Nat. Rev. Gastroenterol. Hepatol.* *12*, 124.
- Guarente, L. (2007). Sirtuins in aging and disease. *Cold Spring Harb. Symp. Quant. Biol.* *72*, 483–488.
- Guo, X., Kimura, A., Azuchi, Y., Akiyama, G., Noro, T., Harada, C., Namekata, K., and Harada, T. (2016). Caloric restriction promotes cell survival in a mouse model of normal tension glaucoma. *Sci. Rep.* *6*, 33950.
- Gupta, S.C., Patchva, S., and Aggarwal, B.B. (2013). Therapeutic roles of curcumin: lessons learned from clinical trials. *AAPS J.* *15*, 195–218.
- Gutiérrez-Salmeán, G., Meaney, E., Lanaspá, M.A., Cicerchi, C., Johnson, R.J., Dugar, S., Taub, P., Ramirez-Sánchez, I., Villarreal, F., Schreiner, G., et al. (2016). A randomized, placebo-controlled, double-blind study on the effects of (-)-epicatechin on the triglyceride/HDLc ratio and cardiometabolic profile of subjects with hypertriglyceridemia: unique in vitro effects. *Int. J. Cardiol.* *223*, 500–506.
- Harrison, D.E., Strong, R., Sharp, Z.D., Nelson, J.F., Astle, C.M., Flurkey, K., Nadon, N.L., Wilkinson, J.E., Frenkel, K., Carter, C.S., et al. (2009). Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* *460*, 392–395.
- Harrison, D.E., Strong, R., Allison, D.B., Ames, B.N., Astle, C.M., Atamna, H., Fernandez, E., Flurkey, K., Javors, M.A., Nadon, N.L., et al. (2014). Acarbose, 17- α -estradiol, and nordihydroguaiaretic acid extend mouse lifespan preferentially in males. *Aging Cell* *13*, 273–282.
- Hawley, S.A., Fullerton, M.D., Ross, F.A., Schertzer, J.D., Chevztzoff, C., Walker, K.J., Peggie, M.W., Zibrova, D., Green, K.A., Mustard, K.J., et al. (2012). The ancient drug salicylate directly activates AMP-activated protein kinase. *Science* *336*, 918–922.
- He, L., Sabet, A., Djedjos, S., Miller, R., Sun, X., Hussain, M.A., Radovick, S., and Wondisford, F.E. (2009). Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. *Cell* *137*, 635–646.
- Heilbronn, L.K., de Jonge, L., Frisard, M.I., DeLany, J.P., Larson-Meyer, D.E., Rood, J., Nguyen, T., Martin, C.K., Volaufova, J., Most, M.M., et al. (2006). Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA* *295*, 1539–1548.
- Herranz, D., Muñoz-Martin, M., Cañamero, M., Mulero, F., Martínez-Pastor, B., Fernández-Capetillo, O., and Serrano, M. (2010). Sirt1 improves healthy ageing and protects from metabolic syndrome-associated cancer. *Nat. Commun.* *1*, 3.
- Hu, J., Webster, D., Cao, J., and Shao, A. (2018). The safety of green tea and green tea extract consumption in adults – results of a systematic review. *Regul. Toxicol. Pharmacol.* *95*, 412–433.
- Huang, R., Xu, Y., Wan, W., Shou, X., Qian, J., You, Z., Liu, B., Chang, C., Zhou, T., Lippincott-Schwartz, J., et al. (2015). Deacetylation of nuclear LC3 drives autophagy initiation under starvation. *Mol. Cell* *57*, 456–466.
- Hubbard, B.P., and Sinclair, D.A. (2014). Small molecule SIRT1 activators for the treatment of aging and age-related diseases. *Trends Pharmacol. Sci.* *35*, 146–154.

- Il'yasova, D., Fontana, L., Bhapkar, M., Pieper, C.F., Spasojevic, I., Redman, L.M., Das, S.K., Huffman, K.M., and Kraus, W.E.; CALERIE Study Investigators (2018). Effects of 2 years of caloric restriction on oxidative status assessed by urinary F2-isoprostanes: the CALERIE 2 randomized clinical trial. *Aging Cell* 17.
- Imenshahidi, M., and Hosseinzadeh, H. (2016). Berberis vulgaris and berberine: an update review. *Phytother. Res.* 30, 1745–1764.
- James, A.M., Hoogewijs, K., Logan, A., Hall, A.R., Ding, S., Fearnley, I.M., and Murphy, M.P. (2017). Non-enzymatic N-acetylation of lysine residues by acetylCoA often occurs via a proximal S-acetylated thiol intermediate sensitive to glyoxalase II. *Cell Rep.* 18, 2105–2112.
- Jung, H.Y., Lee, D., Ryu, H.G., Choi, B.H., Go, Y., Lee, N., Lee, D., Son, H.G., Jeon, J., Kim, S.H., et al. (2017). Myricetin improves endurance capacity and mitochondrial density by activating SIRT1 and PGC-1 α . *Sci. Rep.* 7, 6237.
- Kanfi, Y., Naiman, S., Amir, G., Peshti, V., Zinman, G., Nahum, L., Bar-Joseph, Z., and Cohen, H.Y. (2012). The sirtuin SIRT6 regulates lifespan in male mice. *Nature* 483, 218–221.
- Katsyuba, E., Mottis, A., Zietak, M., De Franco, F.D., Velpen, V., van der Gariani, K., Ryu, D., Cialabrini, L., Matilainen, O., Liscio, P., et al. (2018). De novo NAD⁺ synthesis enhances mitochondrial function and improves health. *Nature* 563, 354–359.
- Khan, S., and Jena, G. (2016). Sodium butyrate reduces insulin-resistance, fat accumulation and dyslipidemia in type-2 diabetic rat: a comparative study with metformin. *Chem. Biol. Interact.* 254, 124–134.
- Kim, Y.C., and Guan, K.L. (2015). mTOR: a pharmacologic target for autophagy regulation. *J. Clin. Invest.* 125, 25–32.
- Kim, J.K., Kim, Y.J., Fillmore, J.J., Chen, Y., Moore, I., Lee, J., Yuan, M., Li, Z.W., Karin, M., Perret, P., et al. (2001). Prevention of fat-induced insulin resistance by salicylate. *J. Clin. Invest.* 108, 437–446.
- Kim, H.S., Montana, V., Jang, H.J., Parpura, V., and Kim, J.A. (2013). Epigallocatechin gallate (EGCG) stimulates autophagy in vascular endothelial cells a potential role for reducing lipid accumulation. *J. Biol. Chem.* 288, 22693–22705.
- Kitani, K., Osawa, T., and Yokozawa, T. (2007). The effects of tetrahydrocurcumin and green tea polyphenol on the survival of male C57BL/6 mice. *Biogerontology* 8, 567–573.
- Kjær, T.N., Ornstrup, M.J., Poulsen, M.M., Stødkilde-Jørgensen, H., Jessen, N., Jørgensen, J.O.L., Richelsen, B., and Pedersen, S.B. (2017). No beneficial effects of resveratrol on the metabolic syndrome: a randomized placebo-controlled clinical trial. *J. Clin. Endocrinol. Metab.* 102, 1642–1651.
- Ko, H., So, Y., Jeon, H., Jeong, M.H., Choi, H.K., Ryu, S.H., Lee, S.W., Yoon, H.G., and Choi, K.C. (2013). TGF- β 1-induced epithelial-mesenchymal transition and acetylation of Smad2 and Smad3 are negatively regulated by EGCG in human A549 lung cancer cells. *Cancer Lett.* 335, 205–213.
- Kolosova, N.G., Vitovtov, A.O., Muraleva, N.A., Akulov, A.E., Stefanova, N.A., and Blagosklonny, M.V. (2013). Rapamycin suppresses brain aging in senescence-accelerated OXYS rats. *Aging* 5, 474–484.
- Kong, Y., Li, K., Fu, T., Wan, C., Zhang, D., Song, H., Zhang, Y., Liu, N., Gan, Z., and Yuan, L. (2016). Quercetin ameliorates A β toxicity in Drosophila AD model by modulating cell cycle-related protein expression. *Oncotarget* 7, 67716–67731.
- Kwon, S., Seok, S., Yau, P., Li, X., Kemper, B., and Kemper, J.K. (2017). Obesity and aging diminish sirtuin 1 (SIRT1)-mediated deacetylation of SIRT3, leading to hyperacetylation and decreased activity and stability of SIRT3. *J. Biol. Chem.* 292, 17312–17323.
- Lagouge, M., Argmann, C., Gerhart-Hines, Z., Meziane, H., Lerin, C., Daussin, F., Messadeq, N., Milne, J., Lambert, P., Elliott, P., et al. (2006). Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1 α . *Cell* 127, 1109–1122.
- Lamming, D.W., Ye, L., Katajisto, P., Goncalves, M.D., Saitoh, M., Stevens, D.M., Davis, J.G., Salmon, A.B., Richardson, A., Ahima, R.S., et al. (2012). Rapamycin-induced insulin resistance is mediated by mTORC2 loss and uncoupled from longevity. *Science* 335, 1638–1643.
- Lasko, L.M., Jakob, C.G., Edalji, R.P., Qiu, W., Montgomery, D., Digiammarino, E.L., Hansen, T.M., Risi, R.M., Frey, R., Manaves, V., et al. (2017). Discovery of a selective catalytic p300/CBP inhibitor that targets lineage-specific tumours. *Nature* 550, 128–132.
- Lee, J.-H., Moon, J.-H., Kim, S.-W., Jeong, J.-K., Nazim, U.M., Lee, Y.-J., Seol, J.-W., and Park, S.-Y. (2015a). EGCG-mediated autophagy flux has a neuroprotection effect via a class III histone deacetylase in primary neuron cells. *Oncotarget* 6, 9701–9717.
- Lee, S.Y., Ku, H.C., Kuo, Y.H., Yang, K.C., Tu, P.C., Chiu, H.L., and Su, M.J. (2015b). Caffeic acid ethanamide prevents cardiac dysfunction through sirtuin dependent cardiac bioenergetics preservation. *J. Biomed. Sci.* 22, 80.
- Lee, W., Lee, S.Y., Son, Y.J., and Yun, J.M. (2015c). Gallic acid decreases inflammatory cytokine secretion Through histone acetyltransferase/histone deacetylase regulation in high glucose-induced human monocytes. *J. Med. Food* 18, 793–801.
- Legeay, S., Rodier, M., Fillon, L., Faure, S., and Clere, N. (2015). Epigallocatechin gallate: a review of its beneficial properties to prevent metabolic syndrome. *Nutrients* 7, 5443–5468.
- Li, Y., and Ding, Y. (2012). Minireview: therapeutic potential of myricetin in diabetes mellitus. *Food Sci. Hum. Wellness* 1, 19–25.
- Li, J., Kim, S.G., and Blenis, J. (2014). Rapamycin: one drug, many effects. *Cell Metab.* 19, 373–379.
- Li, M., Sharma, A., Yin, C., Tan, X., and Xiao, Y. (2017a). Metformin ameliorates hepatic steatosis and improves the induction of autophagy in HFD-induced obese mice. *Mol. Med. Rep.* 16, 680–686.
- Li, X., Yu, W., Qian, X., Xia, Y., Zheng, Y., Lee, J.-H., Li, W., Lyu, J., Rao, G., Zhang, X., et al. (2017b). Nucleus-translocated ACS2 promotes gene transcription for lysosomal biogenesis and autophagy. *Mol. Cell* 66, 684–697.e9.
- Liao, Z.Y., Chen, J.L., Xiao, M.H., Sun, Y., Zhao, Y.X., Pu, D., Lv, A.K., Wang, M.L., Zhou, J., Zhu, S.Y., et al. (2017). The effect of exercise, resveratrol or their combination on sarcopenia in aged rats via regulation of AMPK/Sirt1 pathway. *Exp. Gerontol.* 98, 177–183.
- Lim, J.Y., Oh, M.A., Kim, W.H., Sohn, H.Y., and Park, S.I. (2012). AMP-activated protein kinase inhibits TGF- β -induced fibrogenic responses of hepatic stellate cells by targeting transcriptional coactivator p300. *J. Cell. Physiol.* 227, 1081–1089.
- Liu, Y., Wang, L., Predina, J., Han, R., Beier, U.H., Wang, L.-C.S., Kapoor, V., Bhatti, T.R., Akimova, T., Singhal, S., et al. (2013). Inhibition of p300 impairs Foxp3⁺ T regulatory cell function and promotes antitumor immunity. *Nat. Med.* 19, 1173–1177.
- Liu, C.S., Zheng, Y.R., Zhang, Y.F., and Long, X.Y. (2016). Research progress on berberine with a special focus on its oral bioavailability. *Fitoterapia* 109, 274–282.
- Liu, G., Park, S.H., Imbesi, M., Nathan, W.J., Zou, X., Zhu, Y., Jiang, H., Parisiadou, L., and Gius, D. (2017a). Loss of NAD-dependent protein deacetylase Sirtuin-2 alters mitochondrial protein acetylation and dysregulates mitophagy. *Antioxid. Redox Signal.* 26, 849–863.
- Liu, P.P., Liu, H.H., Sun, S.H., Shi, X.X., Yang, W.C., Su, G.H., and Zhao, J. (2017b). Aspirin alleviates cardiac fibrosis in mice by inhibiting autophagy. *Acta Pharmacol. Sin.* 38, 488–497.
- Madeo, F., Fröhlich, E., and Fröhlich, K.U. (1997). A yeast mutant showing diagnostic markers of early and late apoptosis. *J. Cell Biol.* 139, 729–734.
- Madeo, F., Pietroccola, F., Eisenberg, T., and Kroemer, G. (2014). Caloric restriction mimetics: towards a molecular definition. *Nat. Rev. Drug Discov.* 13, 727.
- Madeo, F., Eisenberg, T., Pietroccola, F., and Kroemer, G. (2018). Spermidine in health and disease. *Science* 359, eaan2788.
- Marin, T.L., Gongol, B., Zhang, F., Martin, M., Johnson, D.A., Xiao, H., Wang, Y., Subramaniam, S., Chien, S., and Shyy, J.Y.-J. (2017). AMPK promotes mitochondrial biogenesis and function by phosphorylating the epigenetic factors DNMT1, RBBP7, and HAT1. *Sci. Signal.* 10, eaaf7478.
- Mariño, G., Pietroccola, F., Eisenberg, T., Kong, Y., Malik, S.A., Andryushkova, A., Schroeder, S., Pendl, T., Harger, A., Niso-Santano, M., et al. (2014). Regulation of autophagy by cytosolic acetyl-coenzyme A. *Mol. Cell* 53, 710–725.

- Martin, C.K., Bhapkar, M., Pittas, A.G., Pieper, C.F., Das, S.K., Williamson, D.A., Scott, T., Redman, L.M., Stein, R., Gilhooly, C.H., et al. (2016). Effect of calorie restriction on mood, quality of life, sleep, and sexual function in healthy nonobese adults: the CALERIE 2 randomized clinical trial. *JAMA Intern. Med.* *176*, 743–752.
- Martin-Montalvo, A., Mercken, E.M., Mitchell, S.J., Palacios, H.H., Mote, P.L., Scheibye-Knudsen, M., Gomes, A.P., Ward, T.M., Minor, R.K., Blouin, M.J., et al. (2013). Metformin improves healthspan and lifespan in mice. *Nat. Commun.* *4*, 2192.
- Matboli, M., Eissa, S., Ibrahim, D., Hegazy, M.G.A., Imam, S.S., and Habib, E.K. (2017). Caffeic acid attenuates diabetic kidney disease via modulation of autophagy in a high-fat diet/streptozotocin-induced diabetic rat. *Sci. Rep.* *7*.
- Mattison, J.A., Wang, M., Bernier, M., Zhang, J., Park, S.S., Maudsley, S., An, S.S., Santhanam, L., Martin, B., Faulkner, S., et al. (2014). Resveratrol prevents high fat/sucrose diet-induced central arterial wall inflammation and stiffening in nonhuman primates. *Cell Metab.* *20*, 183–190.
- Mattison, J.A., Colman, R.J., Beasley, T.M., Allison, D.B., Kemnitz, J.W., Roth, G.S., Ingram, D.K., Weindruch, R., de Cabo, R., and Anderson, R.M. (2017). Caloric restriction improves health and survival of rhesus monkeys. *Nat. Commun.* *8*, 14063.
- Mercken, E.M., Mitchell, S.J., Martin-Montalvo, A., Minor, R.K., Almeida, M., Gomes, A.P., Scheibye-Knudsen, M., Palacios, H.H., Licata, J.J., Zhang, Y., et al. (2014). SIRT1/4 extends survival of male mice on a standard diet and preserves bone and muscle mass. *Aging Cell* *13*, 787–796.
- Mereles, D., and Hunstein, W. (2011). Epigallocatechin-3-gallate (EGCG) for clinical trials: more pitfalls than promises? *Int. J. Mol. Sci.* *12*, 5592–5603.
- Mielgo-Ayuso, J., Barrenechea, L., Alcorta, P., Larrarte, E., Margareto, J., and Labayen, I. (2014). Effects of dietary supplementation with epigallocatechin-3-gallate on weight loss, energy homeostasis, cardiometabolic risk factors and liver function in obese women: randomised, double-blind, placebo-controlled clinical trial. *Br. J. Nutr.* *111*, 1263–1271.
- Miles, S.L., McFarland, M., and Niles, R.M. (2014). Molecular and physiological actions of quercetin: need for clinical trials to assess its benefits in human disease. *Nutr. Rev.* *72*, 720–734.
- Miller, R.A., Harrison, D.E., Astle, C.M., Baur, J.A., Boyd, A.R., de Cabo, R., Fernandez, E., Flurkey, K., Javors, M.A., Nelson, J.F., et al. (2011). Rapamycin, but not resveratrol or simvastatin, extends life span of genetically heterogeneous mice. *J. Gerontol. Ser. A* *66*, 191–201.
- Miller, R.A., Harrison, D.E., Astle, C.M., Fernandez, E., Flurkey, K., Han, M., Javors, M.A., Li, X., Nadon, N.L., Nelson, J.F., et al. (2014). Rapamycin-mediated lifespan increase in mice is dose and sex dependent and metabolically distinct from dietary restriction. *Aging Cell* *13*, 468–477.
- Min, S.W., Chen, X., Tracy, T.E., Li, Y., Zhou, Y., Wang, C., Shirakawa, K., Minami, S.S., Defensor, E., Mok, S.A., et al. (2015). Critical role of acetylation in Tau-mediated neurodegeneration and cognitive deficits. *Nat. Med.* *21*, 1154–1162.
- Minor, R.K., Smith, D.L., Sossong, A.M., Kaushik, S., Poosala, S., Spangler, E.L., Roth, G.S., Lane, M., Allison, D.B., de Cabo, R., et al. (2010). Chronic ingestion of 2-deoxy-d-glucose induces cardiac vacuolization and increases mortality in rats. *Toxicol. Appl. Pharmacol.* *243*, 332–339.
- Minor, R.K., Baur, J.A., Gomes, A.P., Ward, T.M., Csiszar, A., Mercken, E.M., Abdelmohsen, K., Shin, Y.-K., Canto, C., Scheibye-Knudsen, M., et al. (2011). SIRT1/2 improves survival and healthspan of obese mice. *Sci. Rep.* *1*, 70.
- Mitchell, S.J., Bernier, M., Aon, M.A., Cortassa, S., Kim, E.Y., Fang, E.F., Palacios, H.H., Ali, A., Navas-Enamorado, I., Di Francesco, A., et al. (2018). Nicotinamide improves aspects of healthspan, but not lifespan, in mice. *Cell Metab.* *27*, 667–676.e4.
- Morselli, E., Maiuri, M.C., Markaki, M., Megalou, E., Pasparakis, A., Palikaras, K., Criollo, A., Galluzzi, L., Malik, S.A., Vitale, I., et al. (2010). Caloric restriction and resveratrol promote longevity through the Sirtuin-1-dependent induction of autophagy. *Cell Death Dis.* *1*, e10.
- Morselli, E., Mariño, G., Bennetzen, M.V., Eisenberg, T., Megalou, E., Schroeder, S., Cabrera, S., Bénit, P., Rustin, P., Criollo, A., et al. (2011). Spermidine and resveratrol induce autophagy by distinct pathways converging on the acetylproteome. *J. Cell Biol.* *192*, 615–629.
- Moruno, F., Pérez-Jiménez, E., and Knecht, E. (2012). Regulation of autophagy by glucose in mammalian cells. *Cells* *1*, 372–395.
- Most, J., Gilmore, L.A., Smith, S.R., Han, H., Ravussin, E., and Redman, L.M. (2018). Significant improvement in cardiometabolic health in healthy nonobese individuals during caloric restriction-induced weight loss and weight loss maintenance. *Am. J. Physiol. Endocrinol. Metab.* *314*, E396–E405.
- Mu, H.N., Li, Q., Pan, C.S., Liu, Y.Y., Yan, L., Hu, B.H., Sun, K., Chang, X., Zhao, X.R., Fan, J.Y., et al. (2015). Caffeic acid attenuates rat liver reperfusion injury through sirtuin 3-dependent regulation of mitochondrial respiratory chain. *Free Radic. Biol. Med.* *85*, 237–249.
- Murtaza, G., Karim, S., Akram, M.R., Khan, S.A., Azhar, S., Mumtaz, A., and Bin Asad, M.H.H. (2014). Caffeic acid phenethyl ester and therapeutic potentials. *BioMed Res. Int.* *2014*, 145342.
- Nair, V., Sreevalsan, S., Basha, R., Abdelrahim, M., Abudayyeh, A., Rodrigues Hoffman, A., and Safe, S. (2014). Mechanism of metformin-dependent inhibition of mammalian target of rapamycin (mTOR) and Ras activity in pancreatic cancer: role of specificity protein (sp) transcription factors. *J. Biol. Chem.* *289*, 27692–27701.
- Nasri, H., and Rafeian-Kopaei, M. (2014). Metformin: current knowledge. *J. Res. Med. Sci.* *19*, 658–664.
- Navrotskaya, V.V., Oxenkrug, G., Vorobyova, L.I., and Summergrad, P. (2012). Berberine prolongs life span and stimulates locomotor activity of *Drosophila melanogaster*. *Am. J. Plant Sci.* *3*, 1037–1040.
- Netea, M.G., Tack, C.J., Netten, P.M., Lutterman, J.A., and Van der Meer, J.W.M. (2001). The effect of salicylates on insulin sensitivity. *J. Clin. Invest.* *108*, 1723–1724.
- Niu, Y., Na, L., Feng, R., Gong, L., Zhao, Y., Li, Q., Li, Y., and Sun, C. (2013). The phytochemical, EGCG, extends lifespan by reducing liver and kidney function damage and improving age-associated inflammation and oxidative stress in healthy rats. *Aging Cell* *12*, 1041–1049.
- Ogata, T., Ideno, Y., Akai, M., Seichi, A., Hagino, H., Iwaya, T., Doi, T., Yamada, K., Chen, A.Z., Li, Y., et al. (2018). Effects of glucosamine in patients with osteoarthritis of the knee: a systematic review and meta-analysis. *Clin. Rheumatol.* *37*, 2479–2487.
- Onakpoya, I., Hung, S.K., Perry, R., Wider, B., and Ernst, E. (2011). The use of garcinia extract (hydroxycitric acid) as a weight loss supplement: a systematic review and meta-analysis of randomised clinical trials. *J. Obes.* *2011*, 509038.
- Onken, B., and Driscoll, M. (2010). Metformin induces a dietary restriction-like state and the oxidative stress response to extend *C. elegans* healthspan via AMPK, LKB1, and SKN-1. *PLoS One* *5*, e8758.
- Owen, M.R., Doran, E., and Halestrap, A.P. (2000). Evidence that metformin exerts its anti-diabetic effects through inhibition of complex 1 of the mitochondrial respiratory chain. *Biochem. J.* *348*, 607–614.
- Pallauf, K., Duckstein, N., and Rimbach, G. (2017). A literature review of flavonoids and lifespan in model organisms. *Proc. Nutr. Soc.* *76*, 145–162.
- Patrignani, P., and Patrono, C. (2016). Aspirin and cancer. *J. Am. Coll. Cardiol.* *68*, 967–976.
- Peleg, S., Feller, C., Forne, I., Schiller, E., Sévin, D.C., Schauer, T., Regnard, C., Straub, T., Prestel, M., Klima, C., et al. (2016). Life span extension by targeting a link between metabolism and histone acetylation in *Drosophila*. *EMBO Rep.* *17*, 455–469.
- Phan, T.T., Shivu, G.N., Choudhury, A., Abozguia, K., Davies, C., Naidoo, U., Ahmed, I., Yousef, Z., Horowitz, J., and Frenneaux, M. (2009). Multi-centre experience on the use of perhexiline in chronic heart failure and refractory angina: old drug, new hope. *Eur. J. Heart Fail.* *11*, 881–886.
- Pietrocola, F., Mariño, G., Lissa, D., Vacchelli, E., Malik, S.A., Niso-Santano, M., Zamzami, N., Galluzzi, L., Maiuri, M.C., and Kroemer, G. (2012). Pro-autophagic polyphenols reduce the acetylation of cytoplasmic proteins. *Cell Cycle* *11*, 3851–3860.
- Pietrocola, F., Lachkar, S., Enot, D.P., Niso-Santano, M., Bravo-San Pedro, J.M., Sica, V., Izzo, V., Maiuri, M.C., Madeo, F., Mariño, G., et al. (2015). Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ.* *22*, 509–516.

- Pietrocola, F., Pol, J., Vacchelli, E., Rao, S., Enot, D.P., Baracco, E.E., Levesque, S., Castoldi, F., Jacquolot, N., Yamazaki, T., et al. (2016). Caloric restriction mimetics enhance anticancer immunosurveillance. *Cancer Cell* 30, 147–160.
- Pietrocola, F., Demont, Y., Castoldi, F., Enot, D., Durand, S., Semeraro, M., Baracco, E.E., Pol, J., Bravo-San Pedro, J.M., Bordenave, C., et al. (2017). Metabolic effects of fasting on human and mouse blood in vivo. *Autophagy* 13, 567–578.
- Pietrocola, F., Castoldi, F., Markaki, M., Lachkar, S., Chen, G., Enot, D.P., Durand, S., Bossut, N., Tong, M., Malik, S.A., et al. (2018). Aspirin recapitulates features of caloric restriction. *Cell Rep.* 22, 2395–2407.
- Pietsch, K., Saul, N., Menzel, R., Stürzenbaum, S.R., and Steinberg, C.E.W. (2009). Quercetin mediated lifespan extension in *Caenorhabditis elegans* is modulated by age-1, daf-2, sek-1 and unc-43. *Biogerontology* 10, 565–578.
- Price, N.L., Gomes, A.P., Ling, A.J.Y., Duarte, F.V., Martin-Montalvo, A., North, B.J., Agarwal, B., Ye, L., Ramadori, G., Teodoro, J.S., et al. (2012). SIRT1 is required for AMPK activation and the beneficial effects of resveratrol on mitochondrial function. *Cell Metab.* 15, 675–690.
- Proshkina, E., Lashmanova, E., Dobrovol'skaya, E., Zemskaya, N., Kudryavtseva, A., Shaposhnikov, M., and Moskalev, A. (2016). Geroprotective and radioprotective activity of quercetin, (-)-epicatechin, and ibuprofen in *Drosophila melanogaster*. *Front. Pharmacol.* 7, 505.
- Pucciarelli, S., Moreschini, B., Micozzi, D., De Fronzo, G.S., Carpi, F.M., Polzonetti, V., Vincenzetti, S., Mignini, F., and Napolioni, V. (2012). Spermidine and spermine are enriched in whole blood of Nona/centenarians. *Rejuvenation Res.* 15, 590–595.
- Qi, Y., Qiu, Q., Gu, X., Tian, Y., and Zhang, Y. (2016). ATM mediates spermidine-induced mitophagy via PINK1 and Parkin regulation in human fibroblasts. *Sci. Rep.* 6.
- Rahmani, A.H., Alsahty, M.A., Aly, S.M., Khan, M.A., and Aldebari, Y.H. (2018). Role of curcumin in disease prevention and treatment. *Adv. Biomed. Res.* 7, 38.
- Rajman, L., Chwalek, K., and Sinclair, D.A. (2018). Therapeutic potential of NAD-boosting molecules: the in vivo evidence. *Cell Metab.* 27, 529–547.
- Raju, N., Sobieraj-Teague, M., Bosch, J., and Eikelboom, J.W. (2016). Updated meta-analysis of aspirin in primary prevention of cardiovascular disease. *Am. J. Med.* 129, e35–e36.
- Ravussin, E., Redman, L.M., Rochon, J., Das, S.K., Fontana, L., Kraus, W.E., Romashkan, S., Williamson, D.A., Meydani, S.N., Villareal, D.T., et al. (2015). A 2-year randomized controlled trial of human caloric restriction: feasibility and effects on predictors of health span and longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* 70, 1097–1104.
- Redman, L.M., Smith, S.R., Burton, J.H., Martin, C.K., Il'yasova, D., and Ravussin, E. (2018). Metabolic slowing and reduced oxidative damage with sustained caloric restriction support the rate of living and oxidative damage theories of aging. *Cell Metab.* 27, 805–815.e4.
- Ren, J., Yang, L., Zhu, L., Xu, X., Ceylan, A.F., Guo, W., Yang, J., and Zhang, Y. (2017). Akt2 ablation prolongs life span and improves myocardial contractile function with adaptive cardiac remodeling: role of Sirt1-mediated autophagy regulation. *Aging Cell* 16, 976–987.
- Roberts, D.J., Tan-Sah, V.P., Ding, E.Y., Smith, J.M., and Miyamoto, S. (2014). Hexokinase-II positively regulates glucose starvation induced autophagy through TORC1 inhibition. *Mol. Cell* 53, 521–533.
- Rogina, B., Helfand, S.L., and Frankel, S. (2002). Longevity regulation by *Drosophila* Rpd3 deacetylase and caloric restriction. *Science* 298, 1745.
- Rolfe, H.M. (2014). A review of nicotinamide: treatment of skin diseases and potential side effects. *J. Cosmet. Dermatol.* 13, 324–328.
- Romashkan, S.V., Das, S.K., Villareal, D.T., Ravussin, E., Redman, L.M., Rochon, J., Bhapkar, M., and Kraus, W.E.; CALERIE Study Group (2016). Safety of two-year caloric restriction in non-obese healthy individuals. *Oncotarget* 7, 19124–19133.
- Sataranatarajan, K., Ikeno, Y., Bokov, A., Feliars, D., Yalamanchili, H., Lee, H.J., Mariappan, M.M., Tabatabai-Mir, H., Diaz, V., Prasad, S., et al. (2016). Rapamycin increases mortality in db/db mice, a mouse model of type 2 diabetes. *J. Gerontol. A Biol. Sci. Med. Sci.* 71, 850–857.
- Sato, S., Solanas, G., Peixoto, F.O., Bee, L., Symeonidi, A., Schmidt, M.S., Brenner, C., Masri, S., Benitah, S.A., and Sassone-Corsi, P. (2017). Circadian reprogramming in the liver identifies metabolic pathways of aging. *Cell* 170, 664–677.e11.
- Satoh, A., Brace, C.S., Rensing, N., Cliften, P., Wozniak, D.F., Herzog, E.D., Yamada, K.A., and Imai, S.-I. (2013). Sirt1 extends life span and delays aging in mice through the regulation of NK2 homeobox 1 in the DMH and LH. *Cell Metab.* 18, 416–430.
- Saul, N., Pietsch, K., Stürzenbaum, S.R., Menzel, R., and Steinberg, C.E.W. (2011). Diversity of polyphenol action in *Caenorhabditis elegans*: between toxicity and longevity. *J. Nat. Prod.* 74, 1713–1720.
- Schwarz, C., Stekovic, S., Wirth, M., Benson, G., Royer, P., Sigrist, S.J., Pieber, T., Dammbrueck, C., Magnes, C., Eisenberg, T., et al. (2018). Safety and tolerability of spermidine supplementation in mice and older adults with subjective cognitive decline. *Aging* 10, 19–33.
- Schwer, B., Eckersdorff, M., Li, Y., Silva, J.C., Fermin, D., Kurtev, M.V., Gialourakis, C., Comb, M.J., Alt, F.W., and Lombard, D.B. (2009). Calorie restriction alters mitochondrial protein acetylation. *Aging Cell* 8, 604–606.
- Shintani, T., Yamazaki, F., Katoh, T., Umekawa, M., Matahira, Y., Hori, S., Kakizuka, A., Totani, K., Yamamoto, K., and Ashida, H. (2010). Glucosamine induces autophagy via an mTOR-independent pathway. *Biochem. Biophys. Res. Commun.* 391, 1775–1779.
- Shoba, G., Joy, D., Joseph, T., Majeed, M., Rajendran, R., and Srinivas, P.S.S.R. (1998). Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med.* 64, 353–356.
- Shukla, S., Sharma, A., Pandey, V.K., Raisuddin, S., and Kakkar, P. (2016). Concurrent acetylation of FoxO1/3a and p53 due to sirtuin inhibition elicit Bim/PUMA mediated mitochondrial dysfunction and apoptosis in berberine-treated HepG2 cells. *Toxicol. Appl. Pharmacol.* 291, 70–83.
- Si, H., Fu, Z., Babu, P.V.A., Zhen, W., LeRoith, T., Meaney, M.P., Voelker, K.A., Jia, Z., Grange, R.W., and Liu, D. (2011). Dietary epicatechin promotes survival of obese diabetic mice and *Drosophila melanogaster*123. *J. Nutr.* 141, 1095–1100.
- Soda, K., Kano, Y., Sakuragi, M., Takao, K., Lefor, A., and Konishi, F. (2009). Long-term oral polyamine intake increases blood polyamine concentrations. *J. Nutr. Sci. Vitaminol.* 55, 361–366.
- Sonoda, H., Prachasilchai, W., Kondo, H., Yokota-Ikeda, N., Oshikawa, S., Ito, K., and Ikeda, M. (2010). The protective effect of radicicol against renal ischemia-reperfusion injury in mice. *J. Pharmacol. Sci.* 112, 242–246.
- Spindler, S.R., Mote, P.L., Flegal, J.M., and Teter, B. (2013). Influence on longevity of blueberry, cinnamon, green and black tea, pomegranate, sesame, curcumin, morin, pycnogenol, quercetin, and taxifolin fed iso-calorically to long-lived, F1 hybrid mice. *Rejuvenation Res.* 16, 143–151.
- Steffen, Y., Gruber, C., Schewe, T., and Sies, H. (2008). Mono-O-methylated flavanols and other flavonoids as inhibitors of endothelial NADPH oxidase. *Arch. Biochem. Biophys.* 469, 209–219.
- Strong, R., Miller, R.A., Astle, C.M., Floyd, R.A., Flurkey, K., Hensley, K.L., Javors, M.A., Leeuwenburgh, C., Nelson, J.F., Ongini, E., et al. (2008). Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. *Aging Cell* 7, 641–650.
- Su, H., Yang, F., Wang, Q., Shen, Q., Huang, J., Peng, C., Zhang, Y., Wan, W., Wong, C.C.L., Sun, Q., et al. (2017). VPS34 acetylation controls its lipid kinase activity and the initiation of canonical and non-canonical autophagy. *Mol. Cell* 67, 907–921.e7.
- Sun, T., Li, X., Zhang, P., Chen, W.D., Zhang, H.L., Li, D.D., Deng, R., Qian, X.J., Jiao, L., Ji, J., et al. (2015). Acetylation of Beclin 1 inhibits autophagosome maturation and promotes tumour growth. *Nat. Commun.* 6, 7215.
- Sun, Y., Xia, M., Yan, H., Han, Y., Zhang, F., Hu, Z., Cui, A., Ma, F., Liu, Z., Gong, Q., et al. (2018). Berberine attenuates hepatic steatosis and enhances energy expenditure in mice by inducing autophagy and fibroblast growth factor 21. *Br. J. Pharmacol.* 175, 374–387.
- Surh, Y.J., and Na, H.K. (2016). Therapeutic potential and molecular targets of piceatannol in chronic diseases. *Adv. Exp. Med. Biol.* 928, 185–211.
- Tang, D., Tao, S., Chen, Z., Koliesnik, I.O., Calmes, P.G., Hoerr, V., Han, B., Gebert, N., Zörnig, M., Löffler, B., et al. (2016). Dietary restriction improves

- repopulation but impairs lymphoid differentiation capacity of hematopoietic stem cells in early aging. *J. Exp. Med.* **213**, 535–553.
- Tang, X., Chen, X.F., Wang, N.Y., Wang, X.M., Liang, S.T., Zheng, W., Lu, Y.B., Zhao, X., Hao, D.L., Zhang, Z.Q., et al. (2017). SIRT2 acts as a cardioprotective deacetylase in pathological cardiac hypertrophy. *Circulation* **136**, 2051–2067.
- Timmers, S., Konings, E., Bilet, L., Houtkooper, R.H., van de Weijer, T., Goossens, G.H., Hoeks, J., van der Krieken, S., Ryu, D., Kersten, S., et al. (2011). Calorie restriction-like effects of 30 days of resveratrol (resVida™) supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metab.* **14**, 612–622.
- Tissenbaum, H.A., and Guarente, L. (2001). Increased dosage of a sir-2 gene extends lifespan in *Caenorhabditis elegans*. *Nature* **410**, 227–230.
- Trammell, S.A.J., Schmidt, M.S., Weidemann, B.J., Redpath, P., Jaksch, F., Dellinger, R.W., Li, Z., Abel, E.D., Migaud, M.E., and Brenner, C. (2016). Nicotinamide riboside is uniquely and orally bioavailable in mice and humans. *Nat. Commun.* **7**, 12948.
- Tyzka-Czochara, M., Konieczny, P., and Majka, M. (2017). Caffeic acid expands anti-tumor effect of metformin in human metastatic cervical carcinoma HTB-34 cells: implications of AMPK activation and impairment of fatty acids de novo biosynthesis. *Int. J. Mol. Sci.* **18**, E462.
- Unno, K., Ishikawa, Y., Takabayashi, F., Sasaki, T., Takamori, N., Iguchi, K., and Hoshino, M. (2009). Daily ingestion of green tea catechins from adulthood suppressed brain dysfunction in aged mice. *BioFactors* **34**, 263–271.
- van den Bosch, T., Boichenko, A., Leus, N.G.J., Ourailidou, M.E., Wapenaar, H., Rotili, D., Mai, A., Imhof, A., Bischoff, R., Haisma, H.J., et al. (2016). The histone acetyltransferase p300 inhibitor C646 reduces pro-inflammatory gene expression and inhibits histone deacetylases. *Biochem. Pharmacol.* **102**, 130–140.
- van der Meer, A.J., Scicluna, B.P., Moerland, P.D., Lin, J., Jacobson, E.W., Vlasuk, G.P., and van der Poll, T. (2015). The selective sirtuin 1 activator SRT2104 reduces endotoxin-induced cytokine release and coagulation activation in humans. *Crit. Care Med.* **43**, e199–e202.
- van Deursen, J.M. (2014). The role of senescent cells in ageing. *Nature* **509**, 439–446.
- Vauzour, D., Rodriguez-Mateos, A., Corona, G., Oruna-Concha, M.J., and Spencer, J.P.E. (2010). Polyphenols and human health: prevention of disease and mechanisms of action. *Nutrients* **2**, 1106–1131.
- Vella, S., Penna, I., Longo, L., Poggia, G., Garbati, P., Florio, T., Rossi, F., and Pagano, A. (2015). Perhexiline maleate enhances antitumor efficacy of cisplatin in neuroblastoma by inducing over-expression of NDM29 ncRNA. *Sci. Rep.* **5**, 18144.
- Wagner, A.E., Piegholdt, S., Rabe, D., Baenas, N., Schloesser, A., Eggensdorfer, M., Stocker, A., and Rimbach, G. (2015). Epigallocatechin gallate affects glucose metabolism and increases fitness and lifespan in *Drosophila melanogaster*. *Oncotarget* **6**, 30568–30578.
- Wan, W., You, Z., Xu, Y., Zhou, L., Guan, Z., Peng, C., Wong, C.C.L., Su, H., Zhou, T., Xia, H., et al. (2017). mTORC1 phosphorylates acetyltransferase p300 to regulate autophagy and lipogenesis. *Mol. Cell* **68**, 323–335.e6.
- Wang, C., Zhang, X., Teng, Z., Zhang, T., and Li, Y. (2014). Downregulation of PI3K/Akt/mTOR signaling pathway in curcumin-induced autophagy in APP/PS1 double transgenic mice. *Eur. J. Pharmacol.* **740**, 312–320.
- Wang, A., Huen, S.C., Luan, H.H., Yu, S., Zhang, C., Gallezot, J.D., Booth, C.J., and Medzhitov, R. (2016). Opposing effects of fasting metabolism on tissue tolerance in bacterial and viral inflammation. *Cell* **166**, 1512–1525.e12.
- Wang, H., Liu, C., Mei, X., Cao, Y., Guo, Z., Yuan, Y., Zhao, Z., Song, C., Guo, Y., and Shen, Z. (2017). Berberine attenuated pro-inflammatory factors and protect against neuronal damage via triggering oligodendrocyte autophagy in spinal cord injury. *Oncotarget* **8**, 98312–98321.
- Webster, B.R., Scott, I., Han, K., Li, J.H., Lu, Z., Stevens, M.V., Malide, D., Chen, Y., Samsel, L., Connelly, P.S., et al. (2013). Restricted mitochondrial protein acetylation initiates mitochondrial autophagy. *J. Cell Sci.* **126**, 4843–4849.
- van de Weijer, T., Phielix, E., Bilet, L., Williams, E.G., Ropelle, E.R., Bierwagen, A., Livingstone, R., Nowotny, P., Sparks, L.M., Pagliarlunga, S., et al. (2015). Evidence for a direct effect of the NAD⁺ precursor acipimox on muscle mitochondrial function in humans. *Diabetes* **64**, 1193–1201.
- Willcox, B.J., and Willcox, D.C. (2014). Caloric restriction, caloric restriction mimetics, and healthy aging in Okinawa: controversies and clinical implications. *Curr. Opin. Clin. Nutr. Metab. Care* **17**, 51–58.
- Williams, P.A., Harder, J.M., Foxworth, N.E., Cochran, K.E., Philip, V.M., Porciatti, V., Smithies, O., and John, S.W.M. (2017). Vitamin B3 modulates mitochondrial vulnerability and prevents glaucoma in aged mice. *Science* **355**, 756–760.
- Wirth, M., Benson, G., Schwarz, C., Köbe, T., Grittner, U., Schmitz, D., Sigrist, S.J., Bohlken, J., Stekovic, S., Madeo, F., et al. (2018). The effect of spermidine on memory performance in older adults at risk for dementia: a randomized controlled trial. *Cortex* **109**, 181–188.
- Xiao, K., Jiang, J., Guan, C., Dong, C., Wang, G., Bai, L., Sun, J., Hu, C., and Bai, C. (2013). Curcumin induces autophagy via activating the AMPK signaling pathway in lung adenocarcinoma cells. *J. Pharmacol. Sci.* **123**, 102–109.
- Xie, Z., Lau, K., Eby, B., Lozano, P., He, C., Pennington, B., Li, H., Rath, S., Dong, Y., Tian, R., et al. (2011). Improvement of cardiac functions by chronic metformin treatment is associated with enhanced cardiac autophagy in diabetic OVE26 mice. *Diabetes* **60**, 1770–1778.
- Xu, C., Cai, Y., Fan, P., Bai, B., Chen, J., Deng, H.B., Che, C.M., Xu, A., Vanhoutte, P.M., and Wang, Y. (2015). Calorie restriction prevents metabolic aging caused by abnormal SIRT1 function in adipose tissues. *Diabetes* **64**, 1576–1590.
- Yin, J., Xing, H., and Ye, J. (2008). Efficacy of berberine in patients with Type 2 diabetes mellitus. *Metabolism* **57**, 712–717.
- Yoshino, J., Mills, K.F., Yoon, M.J., and Imai, S. (2011). Nicotinamide mononucleotide, a key NAD(+) intermediate, treats the pathophysiology of diet- and age-induced diabetes in mice. *Cell Metab.* **14**, 528–536.
- Yu, L., Li, Q., Yu, B., Yang, Y., Jin, Z., Duan, W., Zhao, G., Zhai, M., Liu, L., Yi, D., et al. (2016). Berberine attenuates myocardial ischemia/reperfusion injury by reducing oxidative stress and inflammation response: role of silent information Regulator 1. *Oxid. Med. Cell. Longev.* **2016**, 1689602.
- Zhang, C., Liu, J., Huang, G., Zhao, Y., Yue, X., Wu, H., Li, J., Zhu, J., Shen, Z., Haffty, B.G., et al. (2016a). Cullin3-KLHL25 ubiquitin ligase targets ACLY for degradation to inhibit lipid synthesis and tumor progression. *Genes Dev.* **30**, 1956–1970.
- Zhang, C.S., Li, M., Ma, T., Zong, Y., Cui, J., Feng, J.W., Wu, Y.Q., Lin, S.Y., and Lin, S.C. (2016b). Metformin activates AMPK through the lysosomal pathway. *Cell Metab.* **24**, 521–522.
- Zhang, H., Ryu, D., Wu, Y., Gariani, K., Wang, X., Luan, P., D'Amico, D., Ropelle, E.R., Lutoff, M.P., Aebersold, R., et al. (2016c). NAD⁺ repletion improves mitochondrial and stem cell function and enhances life span in mice. *Science* **352**, 1436–1443.
- Zhang, H., Alsaleh, G., Feltham, J., Sun, Y., Riffelmacher, T., Charles, P., Frau, L., Yu, Z., Mohammed, S., Balabanov, S., et al. (2018). Translational control of TFEB and autophagy via eIF5A rejuvenates B cell immunity. *bioRxiv*. <https://doi.org/10.1101/360503>.
- Zheng, J., Cheng, J., Zheng, S., Feng, Q., and Xiao, X. (2018). Curcumin, A polyphenolic curcuminoid with its protective effects and molecular mechanisms in diabetes and diabetic cardiomyopathy. *Front. Pharmacol.* **9**, 472.
- Zhu, Y., Tchkonja, T., Pirtskhalava, T., Gower, A.C., Ding, H., Giorgadze, N., Palmer, A.K., Ikeno, Y., Hubbard, G.B., Lenburg, M., et al. (2015). The Achilles' heel of senescent cells: from transcriptome to senolytic drugs. *Aging Cell* **14**, 644–658.