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ARTICLES

Apolipoprotein E Isoform Phenotype and LDL Subclass Response to a Reduced-Fat Diet

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Abstract

Abstract We investigated the association of apolipoprotein (apo) E isoform phenotype with lipoprotein response to reduced dietary fat intake in 103 healthy men (apoE3/2, n=10; apoE3/3, n=65; and apoE4/3, 4/4, n=28). In a randomized, crossover design, subjects consumed high-fat (46%) and low-fat (24%) diets for 6 weeks each. High-fat LDL cholesterol differed among phenotypes, with apoE4/3, 4/4 > apoE3/3 > apoE3/2. Reduction of LDL cholesterol on the low-fat diet was greater for apoE4/3, 4/4 than apoE3/3 ($P < .05$). There was no significant change in plasma apoB level within any of the apoE phenotype groups on the low-fat diet. This result, together with measurements of LDL subfraction mass by analytical ultracentrifugation, indicated that the primary basis for the diet-induced reduction in LDL cholesterol was not reduced LDL particle number but rather a shift from large, buoyant, cholesterol-rich LDL

particles (flotation rate, 7 to 12) to smaller, denser LDL particles (flotation rate, 0 to 7). The magnitude of this effect was related to apoE phenotype, with progressively greater reductions in levels of large LDL ($P<.01$) from apoE3/2 to apoE3/3 to apoE4/3, 4/4. These results indicate that reduced dietary fat lowers levels of large, buoyant LDL particles by an apoE-dependent mechanism.

LDL cholesterol apolipoprotein E LDL mass diet fatty acids

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Apolipoprotein (apo) E is a protein associated with triglyceride-rich lipoproteins and HDL. A major physiological role of lipoprotein-bound apoE is as a ligand for LDL and remnant receptors¹ and the LDL receptor-related protein.² Three alleles of the apoE gene, $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$, are responsible for the main apoE isoforms: apoE2, apoE3, and apoE4, with relative allele frequencies of approximately 10%, 75%, and 15%, respectively.³ The three alleles result in three homozygous phenotypes, E2/2, E3/3, and E4/4, and three heterozygous phenotypes, E3/2, E4/3, and E4/2. ApoE phenotype accounts for up to 7% of the interindividual variation in total serum cholesterol in the general population.³ The $\epsilon 4$ allele is associated with higher and the $\epsilon 2$ with lower LDL cholesterol (LDL-C) and apoB than is the $\epsilon 3$ allele.^{3 4 5 6 7 8} The effects of these alleles on apoB, however, vary in different populations.⁹ Recently, the $\epsilon 4$ allele has been associated with elevations in plasma triglycerides and low concentrations of HDL cholesterol (HDL-C).¹⁰ Some reports have indicated that the $\epsilon 4$ allele is also associated with increased risk of coronary artery disease (CAD).^{3 11 12}

ApoE phenotype has been reported to influence changes in total cholesterol and LDL-C induced by low-fat diets, with greater responses in subjects with the apoE4 isoform.^{13 14} However, the magnitude of this effect is variable,^{13 14 15 16 17} and the biological mechanism has not been established. Additionally, LDLs in plasma comprise multiple distinct subclasses differing in size, density, and chemical composition,¹⁸ and the relation of apoE isoforms to concentrations and dietary responsiveness of LDL subclasses has not been investigated.

In the present study, we tested whether measurements of LDL subclasses may define more clearly the role of the apoE phenotype in influencing LDL-C response to alteration in dietary fat intake.

Methods

Subjects

Healthy, nonsmoking, male volunteers over age 20 years¹⁹ were recruited through newspaper and radio announcements, flyers, and direct mail contact. Eligibility criteria for acceptance into the study were as follows: (1) no cardiovascular disease, acute illness, or active chronic disease in the past 5 years; (2) plasma total cholesterol concentration <6.72 mmol/L (260 mg/dL) and triglyceride concentration <5.65 mmol/L (500 mg/dL); (3) resting blood pressure <160/105 mm Hg; (4) body weight not greater than 130% of ideal according to the 1985 Metropolitan Life Insurance Co Tables; (5) no use of medication likely to interfere with lipid metabolism; and (6) no apoE2/2 phenotype. Each participant signed a consent form approved by the Committee for the Protection of Human Subjects at Lawrence Berkeley Laboratory, University of California, and participated in a medical interview. One hundred fifty men volunteered for the study. Twenty-one individuals were ineligible and twenty-four were unavailable for follow-up. There was no differential dropout rate with respect to a particular diet regimen or apoE phenotype. Data from two individuals with E4/2 phenotype were not included in the present analysis. Mean±SD values for age and body mass index (BMI) of the 103 men in the study were 48.8±11.1 years (range, 28.0 to 79.0) and 25.5±3.0 kg/m² (range, 17.1 to 35.1), respectively.

Experimental Design

The participants were first randomly assigned (irrespective of apoE phenotype) to outpatient treatment with either a high-fat (46% of energy) or a low-fat (24% of energy) diet for 6 consecutive weeks each. The subjects then switched to the alternate diet for an additional 6 weeks. A 6-week diet was chosen based on the experience in previous longer term diet studies that lipoprotein changes appear to stabilize between 4 and 6 weeks after subjects begin a diet.^{20 21} Registered dietitians instructed the subjects on the experimental diets by giving them 2-week cycle menus demonstrating number and size of servings. Nutrient compositions for the experimental diets (Table 1) were calculated by using MINNESOTA NUTRITION DATA SYSTEM (version 2.1) software developed by the Nutrition Coordinating Center, University of Minnesota, Minneapolis.^{22 23} The change from the 46% high-fat diet to the 24% low-fat diet was achieved primarily by reducing percentage of calories from saturated fat (18% to 5%) and polyunsaturated fat (13% to 4%). The fat was replaced with carbohydrate (which increased from 38% to 60% of energy), and the carbohydrate calories remained equally distributed between simple and complex sources. There were no significant differences between the diets in total calories, percentage of energy from monounsaturated fat (12%) and total protein (16%), cholesterol (0.030 to 0.036 g/1000 kJ), ratio of polyunsaturated fat to saturated fat (0.7), and dietary fiber (0.96 to 1.20 g/1000 kJ). Registered dietitians instructed the subjects to refrain from alcohol during the study and to keep exercise and body weight constant between the two diets.

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Table 1.

Nutrient Content of Experimental Diets

	High Fat	Low Fat
Energy, kJ	12 017	12 000
Fat, %	46.0	23.9
Saturated, %	18.3	5.4
Monounsaturated, %	12.4	12.3
Polyunsaturated, %	12.5	4.0
Carbohydrate, %	38.6	60.0
Protein, %	16.2	16.1
Cholesterol, g/1000 kJ	0.034	0.030
Polyunsaturated fat/saturated fat	0.69	0.74
Dietary fiber, g/1000 kJ	1.17	1.20

Values are the mean of 2-week cycle menu for a 12 000-kJ energy level.

The staff contacted the subjects weekly to encourage motivation. Subjects measured their body weights daily at home, and the staff adjusted energy intake if necessary to minimize weight variability. The subjects were surveyed for dietary intake (4-day food records of Thursday through Sunday),^{22 23 24} body

weight, and plasma lipids and lipoproteins once at screening and once during the last week of each experimental diet. Daily diet deviation records were also used as a measure of dietary compliance. Although half the subjects had the low-fat diet first, we use the expression “diet-induced change,” for every variable, to mean “low-fat value minus high-fat value,” regardless of the actual order of the diets.

Laboratory Analyses

Lipids, Lipoproteins, and Apolipoproteins

Venous blood samples were collected in tubes containing Na₂-EDTA, 1.4 mg/mL, after the subjects had fasted for 12 to 14 hours. Plasma was prepared within 2 hours of collection, and blood and plasma were kept at 4°C until processed. Plasma total cholesterol and triglycerides were determined by enzymatic procedures on a Gilford Impact 400E analyzer. HDL-C was measured after heparin-manganese precipitation of plasma.²⁵ These measurements were consistently in control as monitored by the Centers for Disease Control and Prevention standardization program. LDL-C was calculated from the formula of Friedewald et al²⁶ unless triglycerides were >4.52 mmol/L (400 mg/dL), in which case LDL-C was measured in the density >1.006 g/mL ultracentrifugal plasma fraction.²⁷ ApoA-I and apoB concentrations in plasma were determined by maximal radial immunodiffusion.^{28 29}

Analytical Ultracentrifugation

Lipoproteins were analyzed by analytical ultracentrifugation, a procedure that provides measurements of lipoprotein mass as a function of Svedberg flotation rate (S_f^0 for $d < 1.063$ g/mL lipoproteins and $F_{1.20}^0$ for $d < 1.21$ g/mL lipoproteins). Mass concentrations were determined for total LDL (S_f^0 0 to 12) and for concentrations of four major LDL subclasses: LDL-I (S_f^0 7 to 12), LDL-II (S_f^0 5 to 7), LDL-III (S_f^0 3 to 5), and LDL-IV (S_f^0 0 to 3).¹⁸ Mass concentrations were also determined for IDL (S_f^0 12 to 20) and VLDL (S_f^0 20 to 400). For LDL, this procedure provides a measurement of peak flotation rate as well as density and diameter of the peak LDL for each subject.³⁰ In addition, mass was determined for total HDL ($F_{1.20}^0$ 0 to 9) and for concentrations of two major HDL subclasses, HDL₂ ($F_{1.20}^0$ 3.5 to 9) and HDL₃ ($F_{1.20}^0$ 0 to 3.5).³⁰

ApoE Phenotype

ApoE isoforms were determined by isoelectric focusing of VLDL apolipoproteins.^{31 32} Isoform phenotypes were designated according to recommended nomenclature.³²

For all laboratory analyses, personnel were blinded as to the subjects' identity and high- or low-fat diet treatment.

Statistics

Lipid, lipoprotein, apolipoprotein, and dietary data are presented for three apoE phenotype groups, E3/2 (n=10), E3/3 (n=65), and E4/3 (n=28). Results are expressed as mean±SEM. Three apoE4/4 homozygotes were combined with the E4/3 group for all calculations since removing these three subjects from the analyses did not substantially change the results. Univariate analyses were by the Mann-Whitney two-sample test and the Kruskal-Wallis test for three-group comparisons. The Wilcoxon signed rank test was used for paired difference analyses. These analyses showed no diet treatment order effect, ie, the changes (low-fat value minus high-fat value) were not significantly related to the actual order of the diets (high fat to low fat, n=48 versus low fat to high fat, n=55) for any of the lipid, lipoprotein, or apolipoprotein variables. Multivariate analyses were by multiple regression. SAS software^{33 34} was used to perform the statistical analyses, and two-sided tests of statistical significance were employed.

Results

Mean age and BMI were not significantly different among apoE phenotype groups (data not shown). There were no significant mean diet-induced changes in body weight within or among the groups throughout the experimental period (data not shown).

Nutrient intake as estimated from the 4-day food records indicated good compliance with the experimental diets. Table 2 shows reported dietary intake on the high-fat and low-fat diets for 102 subjects (one participant in the apoE3/3 group did not provide diet records during the study). During the high-fat diet, the percentage fat intakes for the apoE3/2, 3/3, and 4/3 groups were 45%, 45%, and 46%, respectively, with no significant differences among phenotypes. Similarly, the groups achieved a fat intake on the low-fat diet of 24%, 25%, and 24%, respectively, with no significant group differences. For intakes of other major nutrients, there were no significant differences among apoE phenotypes on either the high- or low-fat diets.

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Table 2.

Reported Dietary Intake (4-Day Record) on the High-Fat and Low-Fat Diets by ApoE Phenotype

	ApoE3/2 (n=10)		ApoE3/3 (n=64)		ApoE4/3 (n=28)	
	High Fat	Low Fat	High Fat	Low Fat	High Fat	Low Fat
Energy, kJ	11 975±525	11 838 ±508	12 188±179	11 721±229	11 429±358	11 375 ±413

	ApoE3/2 (n=10)		ApoE3/3 (n=64)		ApoE4/3 (n=28)	
	High Fat	Low Fat	High Fat	Low Fat	High Fat	Low Fat
Fat, %	45.2±0.7	24.2±0.6	45.3±0.3	24.6 ±0.5	45.6±0.4	24.0±0.5
Saturated, %	18.7 ±0.3	5.9±0.2	18.2±0.2	6.1±0.2	18.5±0.2	5.9 ±0.1
Monounsaturated, %	12.2±0.2	12.1±0.4	12.6 ±0.1	11.7±0.2	12.4±0.2	11.7±0.2
Polyunsaturated, %	11.5±0.7	3.9±0.2	11.6±0.2	4.4±0.1	11.9 ±0.3	4.0±0.1
Carbohydrate, %	39.2±0.7	59.7 ±0.7	39.0±0.4	58.5±0.4	38.5±0.4	59.3±0.5
Protein, %	16.2±0.1	16.0±0.2	16.3±0.1	16.7 ±0.2	16.5±0.1	16.5±0.5
Cholesterol, g/1000 kJ	0.038 ±0.002	0.031±0.003	0.037±0.001	0.033±0.001	0.037 ±0.001	0.033±0.001
Polyunsaturated fat/ saturated fat	0.62±0.04	0.67±0.02	0.64±0.01	0.73±0.02	0.64 ±0.02	0.69±0.02
Dietary fiber, g/1000 kJ	1.09 ±0.07	1.16±0.06	1.12±0.02	1.18±0.03	1.10 ±0.05	1.21±0.04

Apo indicates apolipoprotein. There were no significant between-group differences.

Effects of Diet and ApoE Phenotype on Plasma Lipid, Lipoprotein Cholesterol, and Apolipoprotein Concentrations

Table 3 shows plasma concentration of lipids, lipoprotein cholesterol, and apolipoproteins on the high- and low-fat diets by apoE phenotype group. After the high-fat diet, there were no significant differences in triglycerides, total cholesterol, HDL-C, or apoA-I among phenotypes. However, LDL-C ($P<.01$) and apoB ($P<.05$) were significantly different among phenotypes, with progressive increases from apoE3/2 to apoE3/3 to apoE4/3. On the low-fat diet, differences in LDL-C and apoB among apoE phenotypes showed a similar trend to that on the high-fat diet, with progressive increases from apoE3/2 to apoE3/3 to apoE4/3 ($P<.05$).

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Table 3.

Plasma Lipid, Lipoprotein Cholesterol, and Apolipoprotein Concentrations by ApoE Phenotype on High-Fat and Low-Fat Diets After Each Diet Sequence

	ApoE3/2 Phenotype (n=10)			ApoE3/3 Phenotype (n=65)			ApoE4/3, 4/4 Phenotype (n=28)		
	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference
Triglycerides, mmol/L	1.47 ±0.25	2.23±0.47	0.76 ±0.41	1.12±0.07	1.58 ±0.10	0.46 ±0.08 ⁴	1.04 ±0.08	1.37±0.12	0.33 ±0.09
Cholesterol, mmol/L									
Total	5.09 ±0.28	4.94±0.31	-0.15±0.16	5.40±0.12	5.06 ±0.12	-0.34±0.07 ⁴	5.78±0.18	5.15±0.16	-0.63 ±0.10 ⁴
LDL	3.06±0.18	2.75±0.21	-0.31 ±0.13 ²	3.65±0.11	3.26±0.12	-0.39±0.06 ⁴	4.04 ±0.16	3.46±0.14	-0.58±0.11

	ApoE3/2 Phenotype (n=10)			ApoE3/3 Phenotype (n=65)			ApoE4/3, 4/4 Phenotype (n=28)		
	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference
HDL	1.36 ±0.12	1.18±0.07	-0.18±0.10	1.25±0.03	1.07 ±0.03	-0.17±0.02 ⁴	1.26±0.05	1.07±0.04	-0.20 ±0.03 ⁴
ApoB, µmol/L	1.69±0.12	1.67 ±0.10	-0.02±0.05	1.99±0.06	1.97±0.05	-0.01 ±0.03	2.15±0.08	2.11±0.09	-0.04±0.03
ApoA-I, µmol/L	46.0±2.57	44.5±2.07	-1.50 ±1.50	44.8±0.75	40.6±0.54	-4.14±0.54 ⁴	44.8 ±1.11	40.5±1.32	-4.29±0.91

Apo indicates apolipoprotein. To convert values to milligrams per deciliter, multiply by 88.57 for triglycerides, 38.67 for cholesterol, 55.0 for apoB, and 2.80 for apoA-I. Values are mean±SEM. Significance of differences between apoE phenotype groups on both high- and low-fat diets are indicated in text.

1 Comparison of diet-induced differences among phenotype groups by Kruskal-Wallis test.

2 $P < .05$,

3 $P < .001$,

4 $P < .0001$ for low- vs high-fat diet difference.

Table 3[¶] also shows changes in plasma lipids, lipoprotein cholesterol, and apolipoproteins B and A-I on low-fat minus high-fat diets. Triglycerides were increased on the low-fat diet, and total cholesterol, HDL-C, and apoA-I were decreased in all apoE phenotype groups; these changes reached significance only for the apoE3/3 and apoE4/3 groups. ApoB levels did not change significantly in any group after changing to the low-fat diet. Diet-induced decreases in total cholesterol were significantly ($P = .02$) different between phenotypes, with progressively greater reductions from apoE3/2 (-0.15 ± 0.16 mmol/L) to

apoE3/3 (-0.34 ± 0.07 mmol/L) to apoE4/3 (-0.63 ± 0.10 mmol/L). Diet-induced decreases in LDL-C were significant in all groups, but the differences among phenotypes for three-group comparisons did not reach significance ($P=.11$). Two-group comparisons showed that the decrease in LDL-C in apoE4/3 (-0.58 ± 0.10 mmol/L) was significantly ($P=.05$) greater than in apoE3/3 (-0.39 ± 0.06 mmol/L). However, after adjustment for group differences in high-fat LDL-C or LDL total mass (S_f^0 0 to 20), apoE4/3 no longer significantly predicted change in LDL-C. There were no significant differences in diet-induced changes in triglycerides, HDL-C, apoB, or apoA-I among the apoE phenotypes.

Dietary Changes in Lipoprotein Mass Concentrations by ApoE Phenotype

Table 4^u shows plasma lipoprotein mass concentrations after the high- and low-fat diets by apoE phenotype group. After the high-fat diet, there were no significant differences between the groups for mass of VLDL, IDL, LDL-III, LDL-IV, HDL₂, or HDL₃. There were significant ($P<.05$) differences among phenotypes, however, for mass of LDL-I and LDL-II, with progressive increases from apoE3/2 to apoE3/3 to apoE4/3 for both LDL-I and LDL-II. After the low-fat diet, the differences among the groups remained significant ($P<.05$) for LDL-II.

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Table 4.

Plasma Lipoprotein Mass Concentrations by ApoE Phenotype on High-Fat and Low-Fat Diets

	ApoE3/2 (n=10)			ApoE3/3 (n=65)			ApoE4/3 (n=28)		
	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference
VLDL, S_f^0 20-400	119.5 \pm 30.0	203.2 \pm 50.2	83.7 \pm 7.0	77.1 \pm 7.3	125.1 \pm 10.1	48.0 \pm 8.3 ⁵	60.9 \pm 9.0	101.7 \pm 11.0	40.8 \pm 10.5 ⁴

	ApoE3/2 (n=10)			ApoE3/3 (n=65)			ApoE4/3 (n=28)		
	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference
IDL, S _f ^o 12-20	39.1±4.9	39.3±4.3	0.2 ±3.4	33.0±2.0	33.2±2.1	0.2±1.8	31.4±3.3	32.2 ±3.1	0.8±2.6
LDL-I, S _f ^o 7-12	97.8±7.8	77.3±7.5	-20.6±7.0 ²	133.4 ±5.9	99.8±5.7	-33.6±4.3 ⁵	138.4±8.5	82.8 ±4.2	-55.6±6.8 ⁵
LDL-II, S _f ^o 5-7	86.0±10.4	81.0±11.1	-5.1±9.7	123.3 ±4.5	108.7±4.4	-14.5±3.9 ⁴	137.1±6.8	114.7 ±6.1	-22.4±6.5 ⁴
LDL-III, S _f ^o 3-5	55.9±13.0	66.8±10.8	10.9±7.8	60.3±4.9	78.6 ±5.0	18.3±4.1 ⁵	62.4±6.5	91.1±7.5	28.7 ±6.6 ⁴
LDL-IV, S _f ^o 0-3	10.7 ±3.3	18.2±5.1	7.5±5.1	10.9±1.4	17.4±2.0	6.4 ±1.4 ⁵	11.8±1.5	19.1±2.7	7.3±2.4 ³
HDL ₂ , F _{1.20} ^o 3.5-9	43.5 ±13.6	27.1±8.0	-16.5±12.9	36.0±4.2	23.6 ±2.8	-12.4±2.5 ⁵	35.3±5.2	24.4±4.6	-10.9 ±3.4 ³

	ApoE3/2 (n=10)			ApoE3/3 (n=65)			ApoE4/3 (n=28)		
	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference	High Fat	Low Fat	Difference
HDL ₃ , F _{1.20} ^o 0-3.5	207.8±13.6	204.4±14.3	-3.3±7.7	190.7 ±4.2	179.8±3.4	-10.8±3.4 ⁴	186.1±5.1	179.4 ±5.9	-6.7±5.6

Apo indicates apolipoprotein; S_f^o and F_{1.20}^o, peak flotation rate. Values are mean±SEM and are given in milligrams per deciliter. Significance of differences between apoE phenotype groups on both high- and low-fat diets are indicated in the text.

1 Comparison of diet-induced differences among phenotype groups by Kruskal-Wallis test.

2 $P < .05$,

3 $P < .01$,

4 $P < .001$,

5 $P < .0001$ for low- vs high-fat diet difference.

Table 4[¶] also shows diet-induced changes in plasma lipoprotein mass concentrations by apoE phenotype. VLDL mass increased in all groups but reached significance only for the apoE3/3 and apoE4/3 groups. There were no diet-induced differences in IDL within phenotypes. Decreases in mass of LDL-I were significant in all groups and were significantly ($P = .01$) related to apoE phenotype, with progressively greater reductions from apoE3/2 (-20.6 ± 7.0 mg/dL) to apoE3/3 (-33.6 ± 4.3 mg/dL) to apoE4/3 (-55.6 ± 6.8 mg/dL). Two-group comparisons showed that the reduction in LDL-I in apoE4/3 was significantly ($P = .02$) greater than apoE3/3 and significantly ($P < .01$) greater than apoE3/2. Within the LDL-I flotation range, the greatest differences in response between apoE4/3 and apoE3/3 was found for S_f^o 7 to 8 (-27.1 ± 3.1 versus -15.4 ± 2.1 mg/dL, respectively) ($P < .01$). Baseline LDL-C level predicts diet-induced changes in LDL-C,³⁵ and after adjustment for differences in high-fat LDL-C among apoE phenotypes, apoE4/3 was still significantly ($P < .05$) associated with greater

reductions in mass of larger LDL after the low-fat diet (data not shown). Mass of LDL-II, HDL₂, and HDL₃ decreased, and LDL-III and LDL-IV increased in all groups, with significant changes in the apoE3/3 and apoE4/3 groups. There was a trend toward greater decreases in LDL-II and increases in LDL-III from apoE3/2 to apoE3/3 to apoE4/3, but these group differences did not reach significance. Within the LDL-II flotation range, the differences in response between apoE4/3 and apoE3/3 for S_f^o 6 to 7 (−21.8±3.6 versus −13.7±2.4 mg/dL, respectively) were marginally significant (*P*=.08). There were no significant group differences for changes in mass of IDL, LDL-IV, HDL₂, or HDL₃.

Discussion

Differences in dietary lipoprotein response have frequently been examined in relation to apoE genetic polymorphism.^{13 14 15 17 36 37 38 39} Some reports have indicated that the apoE4 isoform is associated with higher LDL-C and a greater response of LDL-C to dietary lipids^{13 14 15 36 37} than are the other isoforms. The effect of apoE phenotype on the variation of LDL-C that has been observed in association with a high-fat diet,^{6 7 40} however, has been reported to disappear with a low-fat diet,¹⁶ a result that was not confirmed in the present study. Other studies report that apoE phenotype is not associated with lipid response to dietary intervention.^{17 38 39 41} There are important differences among these studies that could account for such discrepant results. First, study groups differed in gender, age, and baseline lipid levels. Second, the dietary intervention protocols were not consistent among studies. Finally, most of the studies had relatively small numbers of subjects, and the nonsignificant effect of apoE phenotype on dietary lipid response may be due to lack of statistical power.¹⁰ Those studies that showed a significant apoE effect evaluated comparatively larger sample sizes.^{13 14 15}

Using analyses of LDL heterogeneity, the present study identified a strong relation between apoE phenotype and LDL response to reduced dietary fat intake. ApoE4/3 was significantly related to greater decreases in mass of larger, more buoyant LDL particles (S_f^o >7) but not to changes in smaller, more dense LDL or IDL. Previous discrepant results^{13 14 15 16 17 36 37 38 39 41} on the association of apoE phenotype with differences in dietary LDL-C response may depend on whether cholesterol is transported predominantly in larger or smaller LDL particles.

The mechanism by which apoE polymorphism influences LDL levels is still uncertain but may involve effects of apoE on the catabolism of triglyceride-rich particles.^{42 43 44 45 46} There is also evidence for differential distribution of apoE isoforms among lipoprotein particles. ApoE4 has a greater association with triglyceride-rich lipoproteins than apoE3 and apoE2.^{47 48} Clearance of chylomicron remnants is more rapid in subjects with apoE4 than apoE3 isoforms.⁴² This, coupled with enhanced cholesterol absorption in subjects with apoE4,⁴⁹ may increase the intrahepatic pool of cholesterol, downregulate hepatic cholesterol synthesis and LDL receptor activity, and consequently elevate plasma LDL levels.⁴⁷ ApoE3 and apoE4 bind normally to the LDL receptor, but the reduced binding affinity of apoE2⁵⁰ may upregulate LDL receptor activity and increase LDL clearance. Thus, enhanced uptake of apoE4-containing remnant particles along with the reduced receptor binding of apoE2 could contribute to receptor-mediated differences in the effect of apoE alleles on LDL response to reduced dietary lipid.

It is also possible that differential apoE content of larger LDL or its precursors is responsible for the differential effects on LDL particles.^{51 52 53} ApoE-enriched larger LDL particles could have a greater affinity for binding to LDL receptors.⁵⁴ Low-fat diets are known to increase LDL receptor activity, and apoE-containing lipoproteins have a greater affinity for the LDL receptor than do apoB-100-containing lipoproteins.⁵⁴ Thus, larger LDL particles containing apoE may be more rapidly catabolized.⁵⁵ Since clearance rates are faster for apoE4- than apoE3-containing particles,⁴² this would result in a greater decrease in larger LDL particles in subjects with apoE4 phenotypes.

In the present study, reduction in dietary fat resulted in decreases in LDL-C levels without a change in plasma apoB. Since plasma apoB in normotriglyceridemic subjects primarily reflects the number of LDL particles, the results indicate that the predominant mechanism of the reduction in LDL-C with reduced fat intake does not involve a reduction in the number of LDL particles but rather a shift from larger, cholesterol-rich to smaller, cholesterol-poor LDL particles. This shift in LDL particle size was demonstrated by decreases in mass of larger LDL-I and LDL-II particles and concomitant increases in mass of smaller LDL-III and LDL-IV particles, an effect that decreased progressively from apoE4 to apoE3 to apoE2. It is not known whether this change in LDL particle distribution results from enhanced conversion of larger to smaller LDL or from changes in triglyceride-rich lipoprotein precursors that lead to the preferential production of smaller, lipid-depleted LDL. In either case, it is possible that differences in lipid composition of the smaller particles result in decreased apoE binding and consequently a slower clearance rate, with a weaker relation between clearance of these particles and apoE phenotype.⁵⁶

Several studies have shown that LDL heterogeneity is associated with CAD.^{57 58 59 60 61} Either large, buoyant LDL-I^{57 58} or small, dense LDL-III^{59 60 61} particles are frequently found in patients with CAD. However, recent studies have shown that small, dense LDL is potentially more atherogenic than larger LDL by virtue of its increased susceptibility to oxidative modification^{62 63} and its increased promotion of intracellular cholesterol ester accumulation.⁶¹ Reductions in small LDL have been associated with decreased progression of CAD,^{64 65} but the effects of decreases in larger LDL particles on CAD have not been documented. Substantial therapeutic reductions in levels of LDL-I and LDL-II in patients with CAD are not associated with reduced angiographic progression.⁶⁴ Thus, the therapeutic implications of reduction of larger LDL are unclear. In this regard, it is notable that the association of apoE4 as a risk factor for CAD^{3 11} may be independent of LDL-C levels.¹²

Our finding that reductions in HDL-C and apoA-I accompany reductions in LDL-C in all groups of subjects on the low-fat diet is consistent with others' results.^{66 67 68} Decreases in HDL-C seen on low-fat diets may be an adaptive mechanism reflecting decreased flux of HDL cholesteryl ester transport through the HDL metabolic pathway.⁶⁹

The results of this study demonstrate that apoE phenotypes influence the magnitude of LDL-C reduction on low-fat diets by mechanisms that promote a shift from larger, cholesterol-rich to smaller, cholesterol-depleted LDL particles. Our results apply only to reduction in total fat intake, and it is possible that apoE isoforms operate differently in influencing the response to other dietary manipulations designed to lower LDL-C, such as substitution of monounsaturated or polyunsaturated fat for saturated fat or reduction in cholesterol intake. The present results indicate that the relative magnitude of LDL-C reductions induced by a low-fat diet in subjects with differing apoE phenotypes may depend on whether cholesterol is transported predominantly in

larger or smaller LDL particles.

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References

1. Mahley RW. Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science*. 1988;**240**:622-630.
2. Beisiegel U, Weber W, Ihrke G, Herz J, Stanley KK. The LDL-receptor-related protein, LRP, is an apolipoprotein E-binding protein. *Nature*. 1989;**341**:162-164.
3. Davignon J, Gregg RE, Sing CF. Apolipoprotein E polymorphism and atherosclerosis. *Arteriosclerosis*. 1988;**8**:1-21.
4. Sing CF, Davignon J. Role of the apolipoprotein E polymorphism in determining normal plasma lipid and lipoprotein variation. *Am J Hum Genet*. 1985;**37**:268-285.
5. Eto M, Watanabe K, Ishii K. Reciprocal effects and apolipoprotein E alleles ($\epsilon 2$ and $\epsilon 4$) on plasma lipid levels in normolipidemic subjects. *Clin Genet*. 1986;**29**:477-484.
6. Utermann G. Apolipoprotein E polymorphism in health and disease. *Am Heart J*. 1987;**113**:433-440.
7. Boerwinkle E, Utermann G. Simultaneous effects of the apolipoprotein E polymorphism on apolipoprotein E, apolipoprotein B, and cholesterol metabolism. *Am J Hum Genet*. 1988;**42**:104-112.
8. Xhignesse M, Lussier-Cacan S, Sing CF, Kessling AM, Davignon J. Influences of common variants of apolipoprotein E on measures of lipid metabolism in a sample selected for health. *Arterioscler Thromb*. 1991;**11**:1100-1110.
9. Smit M, De Knijff P, Rosseneu M, Bury J, Klasen E, Frants R, Havekes L. Apolipoprotein E polymorphism in the Netherlands and its effect on plasma lipid and apolipoprotein levels. *Hum Genet*. 1988;**80**:287-292.
10. Dallongeville J, Lussier-Cacan S, Davignon J. Modulation of plasma triglyceride levels by apoE phenotype: a meta-analysis. *J Lipid Res*. 1992;**33**:447-454.

11. Hixson JE. Apolipoprotein E polymorphisms affect atherosclerosis in young males: pathobiological determinants of atherosclerosis in youth (PDAY) research group. *Arterioscler Thromb*. 1991;**11**:1237-1244.
12. Eichner JE, Kuller LH, Orchard TJ, Grandits GA, McCallum LM, Ferrell RE, Neaton JD. Relation of apolipoprotein E phenotype to myocardial infarction and mortality from coronary artery disease. *Am J Cardiol*. 1993;**71**:160-165.
13. Tikkanen MJ, Huttunen JK, Ehnholm C, Pietinen P. Apolipoprotein E4 homozygosity predisposes to serum cholesterol elevation during high-fat diet. *Arteriosclerosis*. 1990;**10**:285-288.
14. Mänttääri M, Koskinen P, Ehnholm C, Huttunen JK, Manninen V. Apolipoprotein E polymorphism influences the serum cholesterol response to dietary intervention. *Metabolism*. 1991;**40**:217-221.
15. Cobb MM, Teitelbaum HS, Risch N, Jekel JJ, Ostfield AM. Influence of dietary fat apolipoprotein E phenotype, and sex on plasma lipoprotein levels. *Circulation*. 1992;**86**:849-857.
16. Xu CF, Boerwinkle E, Tikkanen MJ, Huttunen JK, Humphries SE, Talmud PJ. Genetic variation at the apolipoprotein gene loci contribute to response of plasma lipids to dietary change. *Genet Epidemiol*. 1990;**7**:261-275.
17. Savolainen MJ, Rantala M, Kervinen K, Jarvi L, Suvanto K, Rantala T, Kesaniemi YA. Magnitude of dietary effects on plasma cholesterol concentration: role of sex and apolipoprotein E phenotype. *Atherosclerosis*. 1991;**86**:145-152.
18. Krauss RM, Burke DJ. Identification of multiple subclasses of plasma low density lipoproteins in normal humans. *J Lipid Res*. 1982;**23**:97-104.
19. Austin MA, King MC, Vranizan KM, Newman B, Krauss RM. Inheritance of low-density lipoprotein subclass patterns: results of complex segregation analysis. *Am J Hum Genet*. 1988;**43**:838-46.
20. Kuusi T, Ehnholm C, Huttunen JK, Kostianen E, Pietinen P, Leino U, Uusitalo U, Nikkari T, Iacono JM, Puska P. Concentration and composition of serum lipoproteins during a low-fat diet at two levels of polyunsaturated fat. *J Lipid Res*. 1985;**26**:360-367.
21. Ullmann D, Connor WE, Hatcher LF, Connor SL, Flavell DP. Will a high-carbohydrate, low-fat diet lower plasma lipids and lipoproteins without producing hypertriglyceridemia? *Arterioscler Thromb*. 1991;**11**:1059-1067.
22. Feskanich D, Sielaff BH, Chong K, Buzzard IM. Computerized collection and analysis of dietary intake information. *Comput Methods Programs Biomed*. 1989;**30**:47-57.
23. Schakel SF, Sievert YA, Buzzard IM. Sources of data for developing and maintaining a nutrient database. *J Am Diet Assoc*. 1988;**88**:1268-1271.
24. Jackson B, Dujovne CA, DeCoursey S, Beyer P, Brown EF, Hassanein K. Methods to assess relative reliability of diet records: minimum records for monitoring lipid and caloric intake. *J Am Diet Assoc*. 1986;**86**:1531-1535.

25. Warnick GR, Nguyen T, Albers JJ. Comparison of improved precipitation methods for quantification of high density lipoprotein cholesterol. *Clin Chem*. 1985;**31**:217-222.
26. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low density lipoprotein cholesterol in plasma, without the use of preparatory ultracentrifugation. *Clin Chem*. 1972;**18**:499-502.
27. *Manual of Laboratory Operations*. Lipid Research Clinic Program, lipid and lipoprotein analysis. Dept of Health, Education and Welfare publication No. (NIH) 75-628. 1974.
28. Cheung MC, Albers JJ. The measurement of apolipoprotein AI and AII levels in men and women by immunoassay. *J Clin Invest*. 1977;**60**:43-50.
29. Ouchterlony O, Nilsson L-Q. Immunodiffusion and immunoelectrophoresis. In: Weir DM, ed. *Handbook of Experimental Immunology*. Oxford, England: Blackwell Scientific Publications; 1978:19.10-19.13.
30. Lindgren FT, Jensen LC, Hatch FT. The isolation and quantitative analysis of serum lipoproteins. In: Nelson GJ, ed. *Blood Lipids and Lipoproteins: Quantitation, Composition, and Metabolism*. New York, NY: John Wiley & Sons; 1972:181-274.
31. Weisgraber KH, Rall SC Jr, Mahley RW. Human E apoprotein heterogeneity: cysteine-arginine interchanges in the amino acid sequence of the apo-E isoforms. *J Biol Chem*. 1981;**256**:9077-9083.
32. Zannis VI, Breslow JL, Utermann G, Mahley RW, Weisgraber KH, Havel RJ, Goldstein JL, Brown MS, Schonfeld G, Hazzard WR, et al. Proposed nomenclature of apo-E isoproteins, apo-E genotypes, and phenotypes. *J Lipid Res*. 1982;**23**:911-914.
33. SAS Institute Inc. *SAS Users Guide: Basics*, version 5. Cary, NC: SAS Institute Inc; 1985:1290.
34. SAS Institute Inc. *SAS Users Guide: Statistics*, version 5. Cary, NC: SAS Institute Inc; 1985:956.
35. Denke MA, Frantz ID Jr. Response to a cholesterol-lowering diet: efficacy is greater in hypercholesterolemic subjects even after adjustment for regression to the mean. *Am J Med*. 1993;**94**:626-631.
36. Miettinen TA, Kesäniemi YA. Cholesterol absorption: regulation of cholesterol synthesis and elimination and within-population variations of serum cholesterol levels. *Am J Clin Nutr*. 1989;**49**:629-635.
37. Clifton PM, Kestin M, Abbey M, Drysdale M, Nestel PJ. Relationship between sensitivity to dietary fat and dietary cholesterol. *Arteriosclerosis*. 1990;**10**:394-401.
38. Glatz JFC, Demacker PNM, Turner PR, Katan MB. Response of serum cholesterol to dietary cholesterol in relation to apolipoprotein E phenotype. *Nutr Metab Cardiovasc Dis*. 1991;**1**:13-17.
39. Cobb MM, Risch N. Low-density lipoprotein cholesterol responsiveness to diet in normolipidemic subjects. *Metabolism*. 1993;**42**:7-13.
40. Ehnholm C, Lukka M, Kuusi T, Nikkila E, Utermann G. Apolipoprotein E polymorphism in the Finnish population: gene frequencies and relation to lipoprotein concentrations. *J*

Lipid Res. 1986;**27**:227-235.

41. Boerwinkle E, Brown SA, Rohrbach K, Gotto AM Jr, Patsch W. Role of apolipoprotein E and B gene variation in determining response of lipid, lipoprotein, and apolipoprotein levels to increased dietary cholesterol. *Am J Hum Genet.* 1991;**49**:1145-1154.
42. Weintraub MS, Eisenberg S, Breslow JL. Dietary fat clearance in normal subjects is regulated by genetic variation in apolipoprotein. *Eur J Clin Invest.* 1987;**80**:1571-1577.
43. Demant T, Bedford D, Packard CJ, Shepherd J. Influence of apolipoprotein E polymorphism on apolipoprotein B-100 metabolism in normolipemic subjects. *J Clin Invest.* 1991;**88**:1490-1501.
44. Sehayek E, Eisenberg S. Mechanisms of inhibition by apolipoprotein C of apolipoprotein E-dependent cellular metabolism of human triglyceride-rich lipoproteins through the low density lipoprotein receptor pathway. *J Biol Chem.* 1991;**266**:18259-18267.
45. Gylling H, Miettinen TA. Cholesterol absorption and synthesis related to low density lipoprotein metabolism during varying cholesterol intake in men with different apoE phenotypes. *J Lipid Res.* 1992;**33**:1361-1371.
46. Miettinen TA, Gylling H, Vanhanen H, Ollus A. Cholesterol absorption, elimination, and synthesis related to LDL kinetics during varying fat intake in men with different apoprotein E phenotypes. *Arterioscler Thromb.* 1992;**12**:1044-1052.
47. Gregg RE, Zech LA, Schaefer EJ, Stark D, Wilson D, Brewer HB Jr. Abnormal in vivo metabolism of apolipoprotein E4 in humans. *J Clin Invest.* 1986;**78**:815-821.
48. Steinmetz A, Jakobs C, Motzny S, Kaffarnik H. Differential distribution of apolipoprotein E isoforms in human plasma lipoproteins. *Arteriosclerosis.* 1989;**9**:405-411.
49. Kesäniemi YA, Ehnholm C, Miettinen TA. Intestinal cholesterol absorption efficiency in man is related to apoprotein E phenotype. *J Clin Invest.* 1987;**80**:578-581.
50. Weisgraber KH, Innerarity TL, Mahley RW. Abnormal lipoprotein receptor-binding activity of the human apoprotein E due to cysteine-arginine interchange at a single site. *J Biol Chem.* 1982;**257**:2518-2521.
51. Lee DM, Alaupovic P. Apolipoproteins B, C-III and E in two major subpopulations of low-density lipoproteins. *Biochim Biophys Acta.* 1986;**879**:126-133.
52. Marzetta CA, Rudel LL. A species comparison of low density lipoprotein heterogeneity in nonhuman primates fed atherogenic diets. *J Lipid Res.* 1986;**27**:753-762.
53. Stevenson SC, Sawyer JK, Rudel LL. Role of apolipoprotein E on cholesteryl ester-enriched low density lipoprotein particles in coronary artery atherosclerosis of hypercholesterolemic nonhuman primates. *Arterioscler Thromb.* 1992;**12**:28-40.
54. Chappell DA, Fry GL, Waknitz MA, Berns JJ. Evidence for isomerization during binding of apolipoprotein-B100 to low density lipoprotein receptors. *J Biol Chem.* 1992;**267**:270-279.
55. Yamada N, Shames DM, Stoudemire JB, Havel RJ. Metabolism of lipoproteins containing apolipoprotein B-100 in blood plasma of rabbits: heterogeneity related to the presence

of apolipoprotein E. *Proc Natl Acad Sci U S A*. 1986;**83**:3479-3483.

56. Linga V, Leight MA, Curtiss LK, Marcel YL, St Clair RW, Parks JS. Dietary fish oil-induced decrease in low density lipoprotein binding to fibroblasts is mediated by apolipoprotein E. *J Lipid Res*. 1994;**35**:491-500.

57. Patsch W, Ostlund R, Kuisk I, Levy R, Schonfeld G. Characterization of lipoprotein in a kindred with familial hypercholesterolemia. *J Lipid Res*. 1982;**23**:1196-1205.

58. Campos H, Roederer GO, Lussier-Cacan S, Davignon J, Krauss RM. Predominance of large low density lipoprotein particles in normolipidemic patients with coronary artery disease. *Circulation*. 1991;**84**(suppl II):II-119. Abstract.

59. Austin MA, Breslow JL, Hennekens CH, Buring JE, Willett WC, Krauss RM. Low-density lipoprotein subclass patterns and risk of myocardial infarction. *JAMA*. 1988;**260**:1917-1921.

60. Campos H, Genest JJ Jr, Blijlevens E, McNamara JR, Jenner JL, Ordovas JM, Wilson PWF, Schaefer EJ. Low-density lipoprotein particle size and coronary artery disease. *Arterioscler Thromb*. 1992;**12**:187-195.

61. Jaakkola O, Solakivi T, Tertov VV, Orekhov AN, Miettinen TA, Nikkari T. Characteristics of low-density lipoprotein subfractions from patients with coronary artery disease. *Coron Artery Dis*. 1993;**4**:379-385.

62. de Graaf J, Hak-Lemmers HLM, Hectors MPC, Demacker PNM, Hendriks JCM, Stalenhoef AFH. Enhanced susceptibility to in vitro oxidation of the dense low-density lipoprotein subfraction in healthy subjects. *Arteriosclerosis*. 1991;**11**:298-306.

63. Tribble DL, Holl LG, Wood PD, Krauss RM. Variations in oxidative susceptibility among six low density lipoprotein subfractions of differing density and particle size. *Atherosclerosis*. 1992;**93**:189-199.

64. Krauss RM, Miller BD, Fair JM, Haskell WL, Alderman EL, SCRIP Staff. Reduced progression of coronary artery disease with risk factor intervention in patients with LDL subclass pattern B. *Circulation*. 1992;**86**(suppl I):I-63. Abstract.

65. Watts GF, Mandalia S, Brunt JNH, Slavin BM, Coltart DJ, Lewis B. Independent associations between plasma lipoprotein subfraction levels and the course of coronary artery disease in the St. Thomas' Atherosclerosis Regression Study (STARS). *Metabolism*. 1993;**42**:1461-1467.

66. Gonen B, Patsch W, Kuisk I, Schonfeld G. The effect of short-term feeding of a high carbohydrate diet on HDL subclasses in normal subjects. *Metabolism*. 1981;**30**:1125-1129.

67. Brinton EA, Eisenberg S, Breslow JL. A low-fat diet decreases high density lipoprotein (HDL) cholesterol levels by decreasing HDL apolipoprotein transport rates. *J Clin Invest*. 1990;**85**:144-151.

68. Schaefer EJ, Levy RI, Ernst ND, Van Sant FD, Brewer HB Jr. The effects of low cholesterol, high polyunsaturated fat, and low fat diets on plasma lipid and lipoprotein cholesterol levels in normal and hypercholesterolemic subjects. *Am J Clin Nutr*. 1981;**34**:1758-1763.

69. Hayek T, Ito Y, Azrolan N, Verdery RB, Aalto-Setälä K, Walsh A, Breslow JL. Dietary fat increases high density lipoprotein (HDL) levels both by increasing the transport rates and decreasing the fractional catabolic rates of HDL cholesterol ester and apolipoprotein (apo) A-I: presentation of a new animal model and mechanistic studies in human apo A-I transgenic and control mice. *J Clin Invest.* 1993;**91**:1665-1699.

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